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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6:

A61K 39/00, 39/38, 39/12, C12P 21/04, C12N 7/00, 7/01, 7/02, 7/04, 7/06, 7/08, 15/00, 15/09, 15/63, 15/70, 15/74

(11) International Publication Number:

WO 98/09646

(43) International Publication Date:

12 March 1998 (12.03.98)

(21) International Application Number:

PCT/US97/12955

A1

US

(22) International Filing Date:

31 July 1997 (31.07.97)

(30) Priority Data:

08/708,541

5 September 1996 (05.09.96)

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(60) Parent Application or Grant (63) Related by Continuation

US Filed on

08/708,541 (CIP) 5 September 1996 (05.09.96)

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(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

With international search report.

(54) Title: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

(57) Abstract

A system for the generation of live Birnavirus such as infectious bursal disease virus (IBDV), a segmented double-stranded (ds)RNA virus of the Birnavirdae family, using synthetic transcripts derived from cloned DNA has been developed. Independent full-length cDNA clones were constructed which contained the entire coding and non-coding regions of RNA segments A and B of IBDV, respectively. Synthetic RNAs of both segments were produced by in vitro transcription of linearized plasmids with T7 RNA polymerase. Transfection of Vero cells with combined plus-sense transcripts of both segments generated infectious virus as early as 36 hours post-transfection. The development of a reverse genetics system for dsRNA viruses will greatly facilitate studies of the regulation of viral gene expression pathogenesis, and design of a new generation of live and inactivated vaccines.

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A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

Background of the Invention

Infectious bursal disease virus (IBDV), a member of the *Bimaviridae* family, is the causative agent of a highly immunosuppressive disease in young chickens (Kibenge, F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). Infectious bursal disease (IBD) or Gumboro disease is characterized by the destruction of lymphoid follicles in the bursa of Fabricius. In a fully susceptible chicken flock of 3-6 weeks of age the clinical disease causes severe immunosuppression, and is responsible for losses due to impaired growth, decreased feed efficiency, and death. Susceptible chickens less than 3 weeks old do not exhibit outward clinical signs of the disease but have a marked infection characterized by gross lesions of the bursa.

The virus associated with the symptoms of the disease is called infectious bursal disease virus (IBDV). IBDV is a pathogen of major economic importance to the nation and world's poultry industries. It causes severe immunodeficiency in young chickens by destruction of precursors of antibody-production B cells in the bursa of Fabricius. Immunosuppression causes increased susceptibility to other diseases, and interferes with effective vaccination against Newcastle disease, Marek's disease and infectious bronchitis disease viruses.

There are two known serotypes of IBDV. Serotype I viruses are pathogenic to chickens whereas serotype II viruses infect chickens and turkeys. The infection of turkeys is presently of unknown clinical significance.

IBDV belongs to a group of viruses called *Birnaviridae* which includes other bisegmented RNA viruses such as infectious pancreatic necrosis virus (fish), tellina virus and oyster virus (bivalve mollusks) and drosophila X virus (fruit fly). These viruses all contain high molecular weight (MW) double-stranded RNA genomes.

The capsid of the IBDV virion consists of several structural proteins. As many as nine structural proteins have been reported but there is evidence that some of these may have a precursor-product relationship (Kibenge,

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F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). The designation and molecular weights of the viral proteins (VP) are as shown below.

5	Viral Protein	Molecular Weight
· _	VP1	90 kDa
	VP2	41 kDa
	VP3	32 kDa
	VP4	28 kDa
)	VP5	17 kDa

Two segments of double-stranded RNA were identified in the genome of IBDV. The IBDV genome consists of two segments of double-stranded (ds)RNA that vary between 2827 (segment B) to 3261 (segment A) nucleotide base pairs (Mundt, E. et al., Virology, 209, 10-18 (1995)). The larger segment A encodes a polyprotein which is cleaved by autoproteolysis to form mature viral proteins VP2, VP3 and VP4 (Hudson, P.J. et al., Nucleic Acids Res., 14. 5001-5012 (1986)). VP2 and VP3 are the major structural proteins of the virion. VP2 is the major host-protective immunogen of IBDV, and contains the antigenic regions responsible for the induction of neutralizing antibodies (Azad, et al., Virology, 161, 145-152 (1987)). A second open reading frame (ORF), preceding and partially overlapping the polyprotein gene, encodes a protein (VP5) of unknown function that is present in IBDV-infected cells (Mundt, E., et al., J. Gen. Virol., 76, 437-443, (1995)). The smaller segment B encodes VP1, a 90-kDa multifunctional protein with polymerase and capping enzyme activities (Spies, U., et al., Virus Res., 8, 127-140 (1987): Spies, U., et al., J. Gen. Virol., 71, 977-981 (1990)).

It has been demonstrated that the VP2 protein is the major host protective immunogen of IBDV, and that it contains the antigenic region responsible for the induction of neutralizing antibodies. The region containing the neutralization site has been shown to be highly conformation-dependent. The VP3 protein has been considered to be a group-specific antigen because

it is recognized by monoclonal antibodies directed against it from strains of both serotype I and II viruses. The VP4 protein appears to be a virus-coded protease that is involved in the processing of a precursor polyprotein of the VP2, VP3 and VP4 proteins.

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Although the nucleotide sequences for genome segments A and B of various IBDV strains have been published, it was only recently that the complete 5'- and 3'-noncoding sequences of both segments were determined. The 5'-noncoding region of IBDV segments A and B contain a consensus sequence of 32 nucleotides, whereas the 3'-noncoding terminal sequences of both segments are unrelated, but conserved among IBDV strains of the same serotype (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). These terminii might contain sequences important in packaging and in the regulation of IBDV gene expression, as demonstrated for other dsRNA containing viruses such as mammalian and plant reoviruses, and rotaviruses (Anzola, et al., *Proc. Natl. Acad. Sci. USA*, 84, 8301-8305 (1987); Zou, S., et al., *Virology*, 186, 377-388 (1992); Gorziglia, M.I., et al., *Proc. Natl. Acad. Sci. USA*, 89, 5784-5788 (1992)).

In recent years, a number of infectious animal RNA viruses have been generated from cloned cDNA using transcripts produced by DNA-dependent RNA polymerase (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). For example poliovirus, a plus-stranded RNA virus; influenza virus, a segmented negative-stranded RNA virus; rabies virus, a non-segmented negative-stranded RNA virus; all were recovered from cloned cDNAs of their respective genomes (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986); Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990); Schnell, M.J., et al., *EMBO J.*, 13, 4195-4205 (1994)). For reovirus, it was shown that transfection of cells with a combination of SSRNA, dsRNA and *in vitro* translated reovirus products generated infectious reovirus when complemented with a helper virus from a different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). However, to date, there has been no report of a recovered infectious virus of segmented dsRNA genome from synthetic RNAs only.

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Summary of the Invention

This invention relates to the infectious bursal disease virus (IBDV) that is associated with Gumboro disease of young chickens. More particularly, this invention relates to a system for the generation of infectious bursal disease virus (IBDV) using synthetic transcripts derived from cloned cDNA. The present invention will facilitate studies of the regulation of viral gene expression, pathogenesis and design of a new generation of live and inactivated vaccines.

Detailed Description of the Invention

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In an effort to develop a reverse genetics system for IBDV, three independent full-length cDNA clones which contain segment A of serotype I strain D78 or serotype II strain 23/82 and segment B of the serotype I strain P2, respectively, were constructed. Synthetic RNAs of segments A and B were produced by *in vitro* transcription reaction on linearized plasmids with T7 RNA polymerase. Transcripts of these segments, either untreated or treated with DNase or RNase, were evaluated for the generation of infectious virus by transfection of Vero cells.

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The present inventors have demonstrated that synthetic transcripts derived from cloned DNA corresponding to the entire genome of a segmented dsRNA animal virus can give rise to a replicating virus. The recovery of infectious virus after transfecting cells with synthetic plus-sense RNAs derived from cloned cDNA of a virus with a dsRNA genome (IBDV) completes the quest of generating reverse infectious systems for RNA viruses. A number of investigators have generated infectious animal RNA viruses from cloned cDNA (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). Van der Werf et al. were first to generate poliovirus, a plus-stranded RNA virus, using synthetic RNA produced by T7 RNA polymerase on cloned cDNA template (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986)). later, Enami et al. rescued influenza virus, a segmented negative-stranded RNA virus (Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990)); and Schnell et al. generated rabies virus, a non-segmented negative-stranded RNA virus, from cloned cDNAs of their respective genomes (Schnell, M.J., et

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al., *EMBO J.*, 13, 4195-4205 (1994)). Roner et al. developed an infectious system for a segmented dsRNA reovirus by transfecting cells with a combination of synthetic ssRNA, dsRNA, *in vitro* translated reovirus products, and complemented with a helper virus of different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). The resulting virus was discriminated from the helper virus by plaque assay. However, in this system the use of a helper virus was necessary. In contrast, the presently described reverse genetics system of IBDV does not require a helper virus or other viral proteins. Transfection of cells with plus-sense RNAs of both segments was sufficient to generate infectious virus (IBDV). The fate of the additional one or four nucleotides, respectively, transcribed at the 3'-end of segment A was not determined. However, this did not prevent the replication of the viral dsRNA. Similar effects were observed for plus-stranded RNA viruses by different investigators (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)).

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Transfection of plus-sense RNAs of both segments into the same cell was necessary for the successful recovery of IBDV. Transfected RNAs of both segments had to be translated by the cellular translation machinery. The polyprotein of segment A was presumably processed into VP2, VP3 and VP4 proteins which form the viral capsid. The translated protein VP1 of segment B probably acted as a RNA-dependent RNA polymerase and transcribed minus-strands from synthetic plus-strands of both segments, and the reaction products formed dsRNA. Recently, Dobos reported that in vitro transcription by the virion RNA-dependent RNA polymerase of infectious pancreatic necrosis virus (IPNV), a prototype virus of the Bimaviridae family, is primed by VP1 and then proceeds via an asymmetric, semiconservative, stranddisplacement mechanism to synthesize only plus strands during replication of the viral genome (Dobos, P., Virology, 208, 10-25 (1995)). The present system shows that synthesis of minus-strands proceeds on the plus-strands. Whether the resulting transcribed minus-strand RNA serves as a template for the transcription of plus-strands or not remains the subject of further investigation.

To prove that the infectious IBDV contained in the supernatants of transfected cells was indeed derived from the synthetic transcripts, an artificial chimera was generated containing segment A of a serotype II strain and segment B of a serotype I strain. Sequence analysis verified this genome combination. The results also indicate that the terminal sequence motifs described by Mundt and Müller are probably responsible for replication, sorting and packaging of the viral genome (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Presence of serotype-specific terminal sequences obviously does not prevent proper replication of serotype II A segment by the action of the RNA-dependent RNA polymerase VP1 of the serotype I segment B. The ability to create recombinant viruses will greatly help in analyzing the precise function of serotype-specific and serotype-common terminal sequences.

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The recovery of infectious IBDV demonstrates that only the plus-strand RNAs of both segments are sufficient to initiate replication of dsRNA. Thus, the results are in agreement with the general features of reovirus and rotavirus replication where the plus-strand RNAs serve as a template for the synthesis of progeny minus-strands to yield dsRNA (Schonberg, M., et al., *Proc. Natl. Acad. Sci.* Patton, J.T., *Virus Res.*, 6, 217-233 (1986); Chen, D., et al., *J. Virol.*, 68, 7030-7039 (1994)). However, the semiconservative, strand displacement mechanisms proposed by Spies et al. and Dobos could not be excluded (Spies, U., et al., *Virus Res.*, 8, 127-140 (1987); Dobos, P., *Virology*, 208, 10-25 (1995)). The development of a reverse genetics system for IBDV will greatly facilitate future studies of gene expression, pathogenesis, and help in the design of new generations of live and inactivated IBDV vaccines.

As used in the present application, the term "synthetic" as applied to nucleic acids indicates that it is a man made nucleic acid in contrast to a naturally occurring nucleic acid. The term implies no limitation as to the method of manufacture, which can be chemical or biological as long as the method of manufacture involves the intervention of man.

The term "cDNA" is intended to encompass any cDNA containing segments A and B and the 5' and 3' noncoding regions of segments A and B.

The term "infectious" as applied to viruses indicates that the virus has the ability to reproduce. The virus can be pathogenic or nonpathogentic and still be infectious.

The present invention provides a system for the generation of infectious bursal disease virus using synthetic RNA transcripts. This system can be used to study the regulation of viral gene expression, pathogenesis, and for the design of a new generation of live and inactivated IBDV vaccines.

The present invention provides a recombinant vector containing at least one copy of the cDNA according to the present invention. The recombinant vector may also comprise other necessary sequences such as expression control sequences, markers, amplifying genes, signal sequences, promoters, and the like, as is known in the art. Useful vectors for this purpose are plasmids, and viruses such as baculoviruses, herpes virus (HVT) and pox viruses, e.g., fowl pox virus, and the like.

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Also provided herein is a host cell transformed with the recombinant vector of the present invention or a host cell transfected with the synthetic RNA of the present invention. The host cell may be a eukaryotic or a prokaryotic host cell. Suitable examples are *E. coli*, insect cell lines such as Sf-9, chicken embryo fibroblast (CEF) cells, chicken embryo kidney (CEK) cells, African green monkey Vero cells and the like.

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Also part of this invention is an IBDV poultry vaccine comprising a poultry protecting amount of a recombinantly produced virus or portion of a virus, wherein the virus is inactivated or modified such that it is no longer virulent.

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The virus can be inactivated by chemical or physical means. Chemical inactivation can be achieved by treating the virus with, for example, enzymes, formaldehyde, β-propiolactone, ethylene-imine or a derivative thereof, an organic solvent (e.g. halogenated hydrocarbon) and or a detergent. If necessary, the inactivating substance can be neutralized after the virus has been inactivated. Physical inactivation can be carried out by subjecting the viruses to radiation such as UV light, X-radiation, or γ-radiation.

The virus can be attenuated by known methods including serial passage, deleting sequences of nucleic acids and site directed mutagenesis either before or after production of the infectious virus to produce a virus which retains sufficient antigenicity but which has reduced virulence.

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Physiologically acceptable carriers for vaccination of poultry are known in the art and need not be further described herein. In addition to being physiologically acceptable to the poultry the carrier must not interfere with the immunological response elicited by the vaccine and/or with the expression of its polypeptide product.

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Other additives, such as adjuvants and stabilizers, among others, may also be contained in the vaccine in amounts known in the art. Preferably, adjuvants such as aluminum hydroxide, aluminum phosphate, plant and animal oils, and the like, are administered with the vaccine in amounts sufficient to enhance the immune response to the IBDV. The amount of adjuvant added to the vaccine will vary depending on the nature of the adjuvant, generally ranging from about 0.1 to about 100 times the weight of the IBDV, preferably from about 1 to about 10 times the weight of the IBDV.

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The vaccine of the present invention may also contain various stabilizers. Any suitable stabilizer can be used including carbohydrates such as sorbitol, mannitol, starch, sucrose, dextrin, or glucose; proteins such as albumin or casein; and buffers such as alkaline metal phosphate and the like. A stabilizer is particularly advantageous when a dry vaccine preparation is prepared by lyophilization.

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The vaccine can be administered by any suitable known method of inoculating poultry including nasally, ophthalmically, by injection, in drinking water, in the feed, by exposure, and the like. Preferably, the vaccine is administered by mass administration techniques such as by placing the vaccine in drinking water or by spraying the animals' environment. When administered by injection, the vaccines are preferably administered parenterally. Parenteral administration as used herein means administration by intravenous, subcutaneous, intramuscular, or intraperitoneal injection.

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The vaccine of the present invention is administered to poultry to prevent IBD anytime before or after hatching. Preferably, the vaccine is administered prior to the time of birth and after the animal is about 6 weeks of age. Poultry is defined to include but not be limited to chickens, roosters, hens, broilers, roasters, breeders, layers, turkeys and ducks.

The vaccine may be provided in a sterile container in unit form or in other amounts. It is preferably stored frozen, below -20°C, and more preferably below -70°C. It is thawed prior to use, and may be refrozen immediately thereafter. For administration to poultry the recombinantly produced virus may be suspended in a carrier in an amount of about 10⁴ to 10⁷ pfu/ml, and more preferably about 10⁵ to 10⁶ pfu/ml in a carrier such as a saline solution. The inactivated vaccine may contain the antigenic equivalent of 10⁴ to 10⁷ pfu/ml suspended in a carrier. Other carriers may also be utilized as is known in the art. Examples of pharmaceutically acceptable carriers are diluents and inert pharmaceutical carriers known in the art. Preferably, the carrier or diluent is one compatible with the administration of the vaccine by mass administration techniques. However, the carrier or diluent may also be compatible with other administration methods such as injection, eye drops, nose drops, and the like.

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The invention also can be used to produce combination vaccines with the IBDV material. The IBDV material can be combined with antigen material of Newcastle Disease Virus Infectious Bronchitis virus, Reo virus, Adeno virus and/or the Marek virus.

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The foregoing embodiments of the present invention are further described in the following Examples. However, the present invention is not limited by the Examples, and variations will be apparent to those skilled in the art without departing from the scope of the present invention.

Brief Description of the Drawings

Figure 1 is a schematic diagram of cDNA constructs used for synthesis of plus-sense ssRNAs of IBDV with T7 RNA polymerase. Construct pUC19FLAD78 contains the cDNA of segment A of IBDV strain D78 and the recombinant plasmid pUC18FLA23 contains the full-length cDNA of segment

A of IBDV strain 23/82. Segment A of IBDV encodes the polyprotein (VP2-VP4-VP3), and the recently identified VP5 protein. Plasmid pUC18FLBP2 contains the cDNA of segment B of strain P2 which encodes the RNA-dependent RNA polymerase (VP1). Virus specific sequences are underlined and the T7 promoter sequences are italicized. Restriction sites are shown in boldface and identified. The cleavage sites of the linearized plasmids are shown by vertical arrows and the transcription directions are marked by horizontal arrows.

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Figure 2 shows an agarose gel analysis of the transcription reaction products that were used for transfection of Vero cells. Synthetic RNAs transcribed *in vitro* using T7 RNA polymerase and linearized plasmids pUC19FLAD78 (lanes 2, 4 and 6) containing the cDNA of segment A of IBDV strain D78, and pUC18FLBP2 (lanes 1, 3 and 5) containing the cDNA of segment B of strain P2, respectively. After transcription, the reaction mixtures were either treated with DNase (lanes 1 and 2), RNase (lanes 3 and 4) or left untreated (lanes 5 and 6). Two µl of the reaction products were analyzed on 1% agarose gel. Lambda DNA, digested with *Hind* III/EcoR I, was used as markers (lane M).

Figure 3 shows a comparison of nucleotide sequences of cloned RT-PCR fragments from segments A and B of the chimeric IBDV strain 23A/P2B (bold-typed) with known sequences of segments A and B of serotype II strain 23/82 and serotype I strain P2, respectively. Nucleotide identities are marked by a colon.

Figure 4 shows the DNA sequence of pUC18FLA23.

Figure 5 shows the DNA sequence of pUC19FLAD78.

Figure 6 shows the DNA sequence of pUC18FLBP2.

EXAMPLES

Viruses and Cells. Two serotype I strains of IBDV, the attenuated P2 strain from Germany and the vaccine strain D78 (Intervet International), and one serotype II strain, the apathogenic 23/82 strain, were propagated in chicken embryo cells (CEC) and purified (Mundt, E. et al., Virology, 209, 10-18 (1995); Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). Vero cells

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were grown in M199 medium supplemented with 5% fetal calf serum (FCS) and used for transfection experiments. Further propagation of the recovered virus and immunofluorescence studies were carried out in Vero cells (Mundt, E., et al., *J. Gen. Virol.*, 76, 437-443, (1995)). For plaque assay, monolayers of secondary CEC were prepared and used (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA Clones of IBDV genome. Fulllength cDNA clones of IBDV segments A and B were independently prepared. The cDNA clones containing the entire coding region of the RNA segment A of strain D78 were prepared using standard cloning procedures and methods (Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). By comparing the D78 terminal sequences with recently published terminal sequences of other IBDV strains (Mundt, E. et al., Virology, 209, 10-18 (1995)), it was observed that D78 cDNA clones lacked the conserved first 17 and last 10 nucleotides at the 5'- and 3'-ends, respectively. Therefore, to construct a full-length cDNA clone of segment A, two primer pairs (A5'-D78, A5-IPD78 and A3'-IPD78) were synthesized and used for PCR amplification (Table 1). The DNA segments were amplified according to the protocol of the supplier (New England Biolabs) using "Deep Vent Polymerase" (high fidelity thermophilic DNA polymerase). Amplified fragments were cloned into the EcoR I site of a pCRII vector (Invitrogen Corp.) to obtain plasmids pCRD78A5' and pCRD78A3', respectively. Each plasmid was digested with EcoR I and Sal I and the resultant fragments were ligated into EcoR I digested pUC19 to obtain plasmid pUC19FLAD78 (SEQ ID NOS:27 AND 29) which now contains a full-length cDNA copy of segment A encoding all the structural proteins (VP2, VP4 and VP3, SEQ ID NO:30) as well as the non-structural VP5 protein (SEQ ID NO:28) (Fig. 1).

Two primer pairs (A5'-23, A5IP23 and A3'-23, A3-IP23; see Table 1) were used for reverse transcription (RT) of viral genomic dsRNA of strain 23/82 using "SuperScript RT II" (RNA directed DNA polymerase with reduced RNase H activity, GIBCO/BRL). The RT reaction products were purified by phenol/chloroform extraction and ethanol precipitation. To obtain two cDNA

fragments bounded by primer pairs A5'-23, A5-IP23 and A3'-23, A3-IP23, respectively, RT reaction products were amplified by PCR using "Deep Vent polymerase". Both RT and PCR were carried out according to the supplier's protocol. Resulting PCR fragments were blunt-end ligated into *Sma* I cleaved pUC18 vector to obtain pUC23A5' and pUC23A3'. The 3'-end of segment A contained in plasmid pUC23A3' was ligated into the *Hind* III-*Bst*B I cleaved plasmid pUC23A5' to establish the full-length cDNA of segment A of strain 23/82. The resulting plasmid was termed pUC18FLA23 (SEQ ID NOS: 31 AND 33)(Fig. 1) and encodes structural proteins VP2, VP3 and VP4 (SEQ ID NO: 32) and non-structural protein VP5 (SEQ ID NO: 34)

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To obtain cDNA clones of segment B of P2 strain, two primer pairs (B5'-P2, B5-IPP2 and B3'-P2, B3-IPP2) were designed according to the published sequences and used for RT-PCR amplification (see Table 1). Using genomic dsRNA as template, cDNA fragments were synthesized and amplified according to the supplier's protocol (Perkin-Elmer Cetus). Amplified fragments were blunt-end ligated into Sma I cleaved pBS vector (Stratagene) to obtain clones pBSP2B5' and pBSP2B3'. To construct a full-length clone of segment B, the 5'-end fragment of plasmid pBSP2B5' was first subcloned between EcoR I and Pst I sites of pUC18 vector to obtain pUCP2B5'. Then the 3'-end fragment of plasmid pBSP2B3' was inserted between the unique Bal II and Pst I sites of plasmid pUCP2B5' to obtain a full-length plasmid pUC18FLBP2 (SEQ ID NO:25) which encodes the VP1 protein (SEQ ID NO: 26) (Fig. 1). Plasmids pUC18FLBP2, pUC18FLA23 and pUC19FLAD78 were completely sequenced by using the "Sequenase" DNA sequencing system (U.S. Biochem.), and the sequence data were analyzed using either "DNASIS" (Pharmacia) or "PC/Gene" (Intelligenetics) software. The integrity of the full-length constructs was tested by in vitro transcription and translation coupled reticulocyte lysate system using T7 RNA polymerase (Promega).

Transcription and Transfection of Synthetic RNAs. Plasmids pUC19FLAD78, pUC18FLA23 and pUC18FLBP2 were digested with *BsrG* I, *Nsi* I and *Pst* I enzymes (see Fig. 1), respectively, and used as templates for *in vitro* transcription with T7 RNA polymerase (Promega). Briefly, restriction

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enzyme cleavage assays were adjusted to 0.5% SDS and incubated with proteinase K (0.5 mg/ml) for 1 hour at 37°C. The linearized DNA templates (~3 μg) were recovered after ethanol precipitation, and were added separately to a transcription reaction mixture (50 μl) containing 40 mM Tris-HCl (pH 7.9), 10 mM NaCl, 6 mM MgCl₂, 2 mM spermidine, 0.5 mM ATP, CTP and UTP each, 0.1 mM GTP, 0.25 mM cap analog [m7G(5') PPP(5') G], 120 units of "RNasin" (ribonuclease inhibitor), 150 units T7 RNA polymerase (Promega), and incubated at 37°C for 1 hour. Synthetic RNA transcripts were purified by phenol/chloroform extraction and ethanol precipitation. As controls, the transcription products were treated with either DNase or RNase (Promega) before the purification step.

Vero cells were grown to 80% confluence in 60 mm dishes and washed once with phosphate-buffered saline (PBS). Three ml of "OPTI-MEM I" (reduced serum medium containing HEPES buffer, sodium bicarbonate, hypoxanthine, thymidine, sodium pyruvate, L-glutamine, trace elements, growth factors and phenol red; from GIBCO/BRL) were added to the monolayers, and the cells were incubated at 37°C for 1 hour in a CO₂ incubator. Simultaneously, 0.15 ml of "OPTI-MEM I" was incubated with 1.25 (N-[1-(2,3-dioleyloxy)propyl]-N,N,N-"Lipofectin" of reagent μg dioleoylphosphatidylethanolamine, trimethylammonium chloride and GIBCO/BRL) for 45 min. in a polystyrene tube at room temperature. Synthetic RNA transcripts of both segments, resuspended in 0.15 ml of diethyl pyrocarbonate-treated water, were added to the OPTI-MEM-Lipofectinmixture, mixed gently, and incubated on ice for 5 min. After removing the "OPTI-MEM" from the monolayers in 60 mm dishes and replacing with fresh 1.5 ml of "OPTI-MEM", the nucleic acid containing mixture was added dropwise to the Vero cells and swirled gently. After 2 hours of incubation at 37°C, the mixture was replaced with M199 medium [CaCl₂ (annhydrous), Fe(NO₃)₃ 9H₂0, KCl, MgSO₄ (anhydrous), NaCl, NaH₂PO₄H₂O, NaHCO₃, L-Alanine, L-Arginine HCl, L-Aspartic acid, L-Cysteine HCl H2O, L-Cysteine 2HCl, L-Glutamic acid, L-Glutamine, Glycine, L-Histidine HCL H₂O, L-Hydroxyproline, L-Isoleucine, L-Leucine, L-Lysine HCl, L-Methionine, L-Phenylalanine, L-

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Proline, L-Serine, L-Threonine, L-Tryptophan, L-Tyrosine 2Na 2H₂O, L-Valine, Alpha tocopherol PO₄ Na₂, Ascorbic Acid, Biotin, Calciferol, D-Calcium pantothenate, Choline chloride, Folic acid, I-Inositol, Menandione NaHSO₃ 3H₂O, Niacin, Nicotinamide, Para-aminobenzoic acid, Pyridoxine HCl, Riboflavin, Thiamine HCl, Vitamin A Acetate, Adenine SO₄, Adenylic Acid, ATP, Na₂, Cholesterol, 2-Deoxy-D-Ribose, D-Glucose, Glutathione, Guanine HCl, Hypoxanthine Na, Phenol Red Na, Ribose, Sodium Acetate (anhydrous), Thymine, Tween 80, Uracil, and Xanthine Na; from Mediatech, Inc.] containing 5% FCS (without rinsing cells) and the cells were further incubated at 37°C for desired time intervals.

Identification of Generated IBDV. CEC were infected with filtered (0.2 μm) supernatant from Vero cells transfected with transcripts of pUC18FLA23 and pUC18FLP2B. 16 hours post-infection, the whole cell nucleic acids were isolated (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Primers were designed according to the published sequences and RT-PCR fragments were amplified, cloned and sequenced (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Sequence data were analyzed by using "DNASIS" software.

Immunofluorescence. Vero cells, grown on cover slips to 80% confluence, were infected with the supernatants derived from transfected Vero cells (after freeze-thawing) and incubated at 37°C for two days. The cells were then washed, fixed with acetone and treated with polyclonal rabbit anti-IBDV serum. After washing, the cells were treated with fluorescein labeled goat-anti-rabbit antibody (Kirkegaard & Perry Lab.) and examined by fluorescence microscope.

Plaque Assay. Monolayers of secondary CEC, grown in 60 mm dishes, were inoculated with the supernatants derived from transfected Vero cells. After 1 hour of infection, the cells were washed once with PBS and overlayed with 0.8% Agar noble (Difco) containing 10% tryptose phosphate broth, 2% FCS, 0.112% NaHCO₃, 10³ units penicillin, 10³ μg/ml streptomycin, 0.25 μg/ml fungizone, 0.005% neutral red, 0.0015% phenol red. The cells

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were incubated at 37°C for 2 to 3 days until plaques could be observed and counted (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA clones of IBDV Genome. To develop a reverse genetics system for the dsRNA virus IBDV, two independent cDNA clones were constructed that contain segment A of strain D78 and segment B of strain P2 (Fig. 1). Each plasmid encoded either the precursor of structural proteins (VP2, VP4, VP3) and VP5 or only VP1 protein (RNA-dependent RNA polymerase). Plasmid pUC18FLBP2 upon digestion with *Pst* I and transcription *in vitro* by T7 RNA polymerase, would yield RNA containing the correct 5'- and 3'-ends. Whereas, upon digestion with *Bsr*G I and transcription, plasmid pUC19FLAD78 would yield RNA containing the correct 5'-end but with additional four nucleotides at the 3'end. Coupled transcription and translation of the above plasmids in a rabbit reticulocyte system yielded protein products that were correctly processed and comigrated with the marker IBDV proteins after fractionating on SDS-polyacrylamide gel and autoradiography (data not shown).

Transcription, Transfection and Generation of Infectious Virus. Plus-sense transcripts of IBDV segment A and B were synthesized separately in vitro with T7 RNA polymerase using linearized full-length cDNA plasmids as templates (see Fig. 2). Although two species of RNA transcripts were observed for segment B on a neutral gel (lanes 1 and 5), fractionation of these samples on a denaturing gel yielded only one transcript-specific band (data not shown). In order to show that plus-sense RNA transcripts of both segments are needed for the generation of infectious virus, the transcription mixtures were incubated with different nucleases, as shown in Fig. 2. Synthetic RNAs recovered after treating the transcription products with DNase (lanes 1+2), RNase (lanes 3+4) or without treatment (lanes 5+6), were used for the transfection of Vero cells. As mock control, Lipofectin alone was used. Five days post-transfection, cytopathic effect (CPE) was only visible in Vero cells transfected with combined transcripts of untreated or DNase-treated transcription products, but not with RNase-treated transcription mixtures or

mock-transfected control. In addition, no CPE was detected when Vero cells were transfected with RNA of only segment A or B (data not shown). These results demonstrate that replication of IBDV ensued after transfection of Vero cells with plus-sense ssRNAs of both segments of IBDV. To verify that the agent causing the CPE in Vero cells was indeed IBDV, transfected Vero cells were freeze-thawed, and supernatants were clarified by centrifugation, and used to infect CEC or Vero cells. CEC infected with the supernatants derived from Vero transfected cells of untreated or DNase-treated transcription mixtures produced CPE in one day post-inoculation (Table 2). However, no CPE could be detected even after five days in CEC, with the supernatants from transfected Vero cells of RNase-treated transcription mixtures, untreated segment A or B transcription mixtures and mock-transfected Vero cells. Similarly, when Vero cells on cover slips were infected with the same supernatants as described above and examined by immunofluorescence staining after 2 days, only supernatants derived from transfected Vero cells of untreated or DNAse-treated transcription mixtures gave positive immunofluorescence signal (Table 2).

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Recovery of Transfectant Virus. To determine the time point for the recovery of infectious virus, Vero cells were transfected with combined RNA transcripts of segments A and B. At 4, 8, 16, 24, 36 and 48 hours post-transfection, the supernatants were examined for the presence of transfectant virus by infectivity and plaque assays, as shown in Table 3. Our results indicate that the virus could be recovered as early as 36 hours after transfection. Virus titer was 2.3 x 10² pfu/ml which appear to drop for samples obtained later than 48 hours after transfection.

Generation of a Chimeric Virus. To prove that plus-sense ssRNA of both segments of IBDV are sufficient for recovery of infectious virus, a chimeric IBDV was generated. Plasmid pUC18FLA23 containing a full-length sequence of segment A of serotype II strain was linearized by *Nsi* I digestion and ssRNA was synthesized *in vitro* using T7 RNA polymerase. The ssRNA transcript specifies the correct 5'-end but contains one additional residue at the 3'-end (Fig. 1). Vero cells were transfected with ssRNA of segment A of

serotype II strain 23/82 and ssRNA of segment B of serotype I strain P2. Five days after transfection when CPE was evident, the supernatant was clarified (after freeze-thawing) and used to infect CEC. After a second passage in CEC, genomic RNA of the virus was analyzed by RT-PCR and sequencing of the PCR products. Primers for segment A were deigned to specifically amplify only segment A sequences derived from the serotype II strain. Primer for segment B bound to sequences of both serotypes. The amplified fragments were cloned and sequenced. The obtained segment A sequences showed a perfect match with known segment A sequences of serotype II strain 23/82, whereas segment B sequence exhibited complete homology to published segment B sequences of serotype I strain P2 (Fig. 3).

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Table 1. Oligonucleotides Used for the Construction of Full Length cDNA Clones of IBDV Genomic Segments A and B.

Nucleotide Sequence	Orientation	Name	Nucleotide Number
TAATACGACTCACTATAGGATACGATCGGTCTGACCCCGGGGGGGG	÷	A5′-D78	1-31
AGAGAATTC TAA TACGACTCACTA TAGGA TACGATCGGTCTGAC	£	A5′-23	1-48
TGTACAGGGGACCCGCGAACGGATCCAATT	(-)	A3'-D78	3237-3261
CGGCGAATTCATGCATAGGGGACCCGCGAACGGATC	(-)	A3′-23	3242-3261
CGTCGACTACGGGATTCTGG	(-)	A5-IPD78	1711-1730
CAGAGGCAGTACTCCGTCTG	(-)	A5-IP23	1971-1990
А GTCGAC GGGAПСПGСП	(+)	A3-IPD78	1723-1742
GAAGGTGTGCGAGGAC	(±)	A3-IP23	1883-1900
AGAGAATTC TAATACGACTCACTATAGGATACGATGGGTCTGAC	(+)	B5′-P2	1-18
CGATCTGCTGCAGGCCCCCCCGCAGGCGAAGG	(-)	B3'-P2	2807-2827
СТТGAGACTCTTGTTCTCTACTCC	(-)	B5-IPP2	1915-1938
ATACAGCAAAGATCTCGGG	(+)	B3-IPP2	1839-1857

Composition and location of the oligonucleotide primers used for cloning. T7 promoter sequences are marked with italic types, the virus specific sequences are underlined, and the restriction sites marked in boldface. Orientation of the virus specific sequence of the primer is shown for sense (+) and antisense (-). The positions where the primers bind (nucleotide number) are according to the published sequences of P2 strain (2).

Table 2. Generation of Infections IBDV From Synthetic RNAs of Segment A and B.

Material Transfected	CPE	Immunofluoroescence
ssRNA A+B, DNase-treated	+	+
ssRNA A+B, RNase-treated	•	-
ssRNA A+B, untreated	• +	+
ssRNA A, untreated	•	_
ssRNA B, untreated	•	-
Lipofectin only	-	

Vero cells were transfected with synthetic RNAs of segment A and B derived from transcription reactions that were either untreated or treated with DNase or RNase. After 5 days, the supernatants were collected, clarified by centrifugation, and analyzed for the presence of virus. The infectivity of the recovered virus was determined in CEC by the appearance of cytopathic effect (CPE) 1-2 days post-inoculation. The specificity of the recovered virus was determined by immunofluorescence staining of infected Vero cells with rabbit anti-IBDV serum.

Table 3. Recovery of Virus at Various Times Post-Transfection.

Time in hours post-transfection	CPE	Immunofluorescence	pfu/ml
4	-	-	0
8	-	-	0
16	-	-	0
24	-	-	0
36	+	+	2.3 × 10 ²
48	+	+	6.0 × 10 ¹

Vero cells were transfected with synthetic RNAs of segment A and B as described. The infectivity and specificity of the recovered virus was detected by CPE in CEC and immunofluorescence staining in Vero cells, respectively. Monolayers of secondary CEC were used for plaque assay after inoculating the cells with the supernatants derived from transfected Vero cells. Approximate titer of the virus was calculated as plaque forming units per ml (pfu/ml).

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SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: VAKHARIA, Vikram N. MUNDT, Egbert
- (ii) TITLE OF INVENTION: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS
 - (iii) NUMBER OF SEQUENCES: 34
 - (iv) CORRESPONDENCE ADDRESS:
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 - (B) STREET: 655 Fifteenth Street, N. W., Suite 330 G Street Lobby
 - (C) CITY: Washington
 - (D) STATE: DC
 - (E) COUNTRY: USA
 - (F) ZIP: 20005-5701
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: KITTS, Monica C.
 - (B) REGISTRATION NUMBER: 36,105
 - (C) REFERENCE/DOCKET NUMBER: P8172-6002
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 202/638-5000
 - (B) TELEFAX: 202/638-4810
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 46 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: circular
 - (ii) MOLECULE TYPE: cDNA

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:	
GAATTCGGCT TTAATACGAC TCACTATAGG ATACGATCGG TCTGAC	46
(2) INFORMATION FOR SEQ ID NO:2:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 41 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
AATTGGATCC GTTCGCGGGT CCCCTGTACA AAGCCGAATT C	41
(2) INFORMATION FOR SEQ ID NO:3:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
CGGCGAATTC ATGCATAGGG GACCCGCGAA CGGATC	36
(2) INFORMATION FOR SEQ ID NO:4:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 44 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: CDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
GTCAGACCGA TCGTATCCTA TAGTGAGTCG TATTAGAATT CTCT	44
(2) INFORMATION FOR SEQ ID NO:5:	

PCT/US97/12955 WO 98/09646

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 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 33 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
TTGCATGCCT GCAGGGGGCC CCCGCAGGCG AAG	33
(2) INFORMATION FOR SEQ ID NO:6:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
TCGTATCCTA TAGTGAGTCG TATTAGAATT C	31
(2) INFORMATION FOR SEQ ID NO:7:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GGAAGCCTGA GTGAGTTGAC TGACTACAGC TACAACGGGC TGATGTCAGC CACTGCGAAC	60
ATCAACGACA AGATCGGGAA CGTTCTAGTT GGAGAAGGGG TGACTGTTCT CAGTCTACCG	120
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 120 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	

(D) TOPOLOGY: linear

24

27	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GGAAGCCTGA GTGAGTTGAC TGACTACAGC TACAACGGGC TGATGTCAGC CACTGCGAAC	60
ATCAACGACA AGATCGGGAA CGTTCTAGTT GGAGAAGGGG TGACTGTTCT CAGTCTACC	119
(2) INFORMATION FOR SEQ ID NO:9:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
GGAAGCCTGA GTGAACTGAC AGATGTTAGC TACAATGGGT TGATGTCTGC AACAGCCAAC	60
ATCAACGACA AAATTGGGAA CGTCCTAGTA GGGGAAGGGG TCACCGTCCT CAGCTTACCC	120
(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TTTTCAATAG TCCACAGGCG CGAACGAAGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CCAAAGTCTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:11:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 120 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	

(ii) MOLECULE TYPE: DNA

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
TTTTCAACAG TCCACAGGCG CGAAGCACGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CTAAAGTTTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:12:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
TTTTCAACAG TCCACAGGCG CGAAGCACGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CTAAAGTTTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:13:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 48 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TAATACGACT CACTATAGGA TACGATCGGT CTGACCCCGG GGGAGTCA	48
(2) INFORMATION FOR SEQ ID NO:14:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 44 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

AGAGAATTCT AATACGACTC ACTATAGGAT ACGATCGGTC TGAC	44
(2) INFORMATION FOR SEQ ID NO:15:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
TGTACAGGGG ACCCGCGAAC GGATCCAATT	30
(2) INFORMATION FOR SEQ ID NO:16:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
CGGCGAATTC ATGCATAGGG GACCCGCGAA CGGATC	36
(2) INFORMATION FOR SEQ ID NO:17:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
CGTCGACTAC GGGATTCTGG	20
(2) INFORMATION FOR SEQ ID NO:18:	
(i) SEQUENCE CHARACTERISTICS:	

(A) LENGTH: 20 base pairs

27

(0) TYPE: nucleic acid) STRANDEDNESS: single) TOPOLOGY: linear	
(ii) MOL	ECULE TYPE: DNA	
(xi) SEQ	UENCE DESCRIPTION: SEQ ID NO:18:	
CAGAGGCAGT A	CTCCGTCTG	20
(2) INFORMAT	ION FOR SEQ ID NO:19:	
(B (C	UENCE CHARACTERISTICS: 1) LENGTH: 20 base pairs 2) TYPE: nucleic acid 2) STRANDEDNESS: single 3) TOPOLOGY: linear	
(ii) MOL	ECULE TYPE: DNA	
(xi) SEQ	QUENCE DESCRIPTION: SEQ ID NO:19:	
AGTCGACGGG A	TTCTTGCTT	20
(2) INFORMAT	TION FOR SEQ ID NO:20:	
(QUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOI	LECULE TYPE: DNA	
(xi) SE(QUENCE DESCRIPTION: SEQ ID NO:20:	
GAAGGTGTGC (GAGAGGAC	18
(2) INFORMA	TION FOR SEQ ID NO:21:	
() ()	QUENCE CHARACTERISTICS: A) LENGTH: 44 base pairs B) TYPE: nucleic acid C) STRANDEDNESS: single D) TOPOLOGY: linear	
(ii) MO	LECULE TYPE: DNA	
(vi) SE	OUENCE DESCRIPTION: SEQ ID NO:21:	

28

AGAGAATTCT AATACGACTC ACTATAGGAT ACGATGGGTC TGAC	44
(2) INFORMATION FOR SEQ ID NO:22:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 33 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
CGATCTGCTG CAGGGGGCCC CCGCAGGCGA AGG	33
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
CTTGAGACTC TTGTTCTCTA CTCC	24
(2) INFORMATION FOR SEQ ID NO:24:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
ATACAGCAAA GATCTCGGG	19
(2) INFORMATION FOR SEQ ID NO:25:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 2827 base pairs(B) TYPE: nucleic acid	

(C)	STRANDEDNE	ESS:	single
(D)	TOPOLOGY:	circ	cular

(ii) MOLECULE TYPE: cDNA

1	÷	x)	FEATURE:	
1	- 1	χз	PEALURE:	

(ix) FEATURE.

(A) NAME/KEY: CDS

(B) LOCATION: 112..2745

(vi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

(XI) SEQUENCE DESCRIPTION. SEQ IS NOTED.																
GGAI	ACGA	ATG C	GTCT	GACC	C T	TGGG	AGTO	: ACC	TAAE	AAC	GTGG	CTAC	TA G	GGGC	GATAC	60
CCGC	CGC1	rgg (CCGCC	CACGI	T AG	STGGC	TCCT	CTI	CTTC	ATG	ATTO	TGCC	AC C		AGT Ser	117
GAC Asp	ATT Ile	TTC Phe 5	AAC Asn	AGT Ser	CCA Pro	CAG Gln	GCG Ala 10	CGA Arg	AGC Ser	ACG Thr	ATC Ile	TCA Ser 15	GCA Ala	GCG Ala	TTC Phe	165
GGC Gly	ATA Ile 20	AAG Lys	CCT Pro	ACT Thr	GCT Ala	GGA Gly 25	CAA Gln	GAC Asp	GTG Val	GAA Glu	GAA Glu 30	CTC Leu	TTG Leu	ATC Ile	CCT Pro	213
AAA Lys 35	GTT Val	TGG Trp	GTG Val	CCA Pro	CCT Pro 40	GAG Glu	GAT Asp	CCG Pro	CTT Leu	GCC Ala 45	AGC Ser	CCT Pro	AGT Ser	CGA Arg	CTG Leu 50	261
GCA Ala	AAG Lys	TTC Phe	CTC Leu	AGA Arg 55	GAG Glu	AAC Asn	GGC Gly	TAC Tyr	AAA Lys 60	GTT Val	TTG Leu	CAG Gln	CCA Pro	CGG Arg 65	TCT Ser	309
CTG Leu	CCC Pro	GAG Glu	AAT Asn 70	GAG Glu	GAG Glu	TAT Tyr	GAG Glu	ACC Thr 75	GAC Asp	CAA Gln	ATA Ile	CTC Leu	CCA Pro 80	GAC Asp	TTA Leu	357
GCA Ala	TGG Trp	ATG Met 85	CGA Arg	CAG Gln	ATA Ile	GAA Glu	GGG Gly 90	GCT Ala	GTT Val	TTA Leu	AAA Lys	CCC Pro 95	ACT Thr	CTA Leu	TCT Ser	405
CTC Leu	CCT Pro 100	Ile	GGA Gly	GAT Asp	CAG Gln	GAG Glu 105	TAC Tyr	TTC Phe	CCA Pro	AAG Lys	TAC Tyr 110	TAC Tyr	CCA Pro	ACA Thr	CAT His	453
CGC Arg 115	Pro	AGC Ser	AAG Lys	GAG Glu	AAG Lys 120	Pro	AAT Asn	GCG Ala	TAC Tyr	CCG Pro 125	Pro	GAC Asp	ATC Ile	GCA Ala	CTA Leu 130	501

PCT/US97/12955

CTC	AAG Lys	CAG Gln	ATG Met	ATT Ile 135	TAC Tyr	CTG Leu	TTT Phe	CTC Leù	CAG Gln 140	GTT Val	CCA Pro	GAG Glu	GCC Ala	AAC Asn 145	GAG Glu	549
GGC Gly	CTA Leu	AAG Lys	GAT Asp 150	GAA Glu	GTA Val	ACC Thr	CTC Leu	TTG Leu 155	ACC Thr	CAA Gln	AAC Asn	ATA Ile	AGG Arg 160	GAC Asp	AAG Lys	597
						TAC Tyr										645
						GGA Gly 185										693
						AGC Ser										741
						GAG Glu										789
						TTG Leu										837
						CTT Leu										885
						CGC Arg 265										933
						TTT Phe										981
						GGG Gly										1029
		Asp				TTA Leu										1077
	Arg					ACA Thr										1125

TCA Ser	GCT Ala 340	CCA Pro	TCC Ser	CCA Pro	ACA Thr	CAC His 345	CTC Leu	ATG Met	ATC Ile	TCT Ser	ATG Met 350	ATC Ile	ACC Thr	TGG Trp	CCC Pro	1173
GTG Val 355	ATG Met	TCC Ser	AAC Asn	AGC Ser	CCA Pro 360	AAT Asn	AAC Asn	GTG Val	TTG Leu	AAC Asn 365	ATT Ile	GAA Glu	GGG Gly	TGT Cys	CCA Pro 370	1221
TCA Ser	CTC Leu	TAC Tyr	AAA Lys	TTC Phe 375	AAC Asn	CCG Pro	TTC Phe	AGA Arg	GGA Gly 380	GGG Gly	TTG Leu	AAC Asn	AGG Arg	ATC Ile 385	GTC Val	1269
GAG Glu	TGG Trp	ATA Ile	TTG Leu 390	GCC Ala	CCG Pro	GAA Glu	GAA Glu	CCC Pro 395	AAG Lys	GCT Ala	CTT Leu	GTA Val	TAT Tyr 400	GCG Ala	GAC Asp	1317
AAC Asn	ATA Ile	TAC Tyr 405	ATT Ile	GTC Val	CAC His	TCA Ser	AAC Asn 410	ACG Thr	TGG Trp	TAC Tyr	TCA Ser	ATT Ile 415	GAC Asp	CTA Leu	GAG Glu	1365
AAG Lys	GGT Gly 420	GAG Glu	GCA Ala	AAC Asn	TGC Cys	ACT Thr 425	CGC Arg	CAA Gln	CAC His	ATG Met	CAA Gln 430	GCC Ala	GCA Ala	ATG Met	TAC Tyr	1413
TAC Tyr 435	ATA Ile	CTC Leu	ACC Thr	AGA Arg	GGG Gly 440	TGG Trp	TCA Ser	GAC Asp	AAC Asn	GGC Gly 445	GAC Asp	CCA Pro	ATG Met	TTC Phe	AAT Asn 450	1461
CAA Gln	ACA Thr	TGG Trp	GCC Ala	ACC Thr 455	TTT Phe	GCC Ala	ATG Met	AAC Asn	ATT Ile 460	GCC Ala	CCT Pro	GCT Ala	CTA Leu	GTG Val 465	GTG Val	1509
GAC Asp	TCA Ser	TCG Ser	TGC Cys 470	CTG Leu	ATA Ile	ATG Met	AAC Asn	CTG Leu 475	CAA Gln	ATT Ile	AAG Lys	ACC Thr	TAT Tyr 480	GGT Gly	CAA Gln	1557
GGC Gly	AGC Ser	GGG Gly 485	AAT Asn	GCA Ala	GCC Ala	ACG Thr	TTC Phe 490	ATC Ile	AAC Asn	AAC Asn	CAC His	CTC Leu 495	TTG Leu	AGC Ser	ACA Thr	1605
CTA Leu	GTG Val 500	CTT Leu	GAC Asp	CAG Gln	TGG Trp	AAC Asn 505	CTG Leu	ATG Met	AGA Arg	CAG Gln	CCC Pro 510	AGA Arg	CCA Pro	GAC Asp	AGC Ser	1653
GAG Glu 515	GAG Glu	TTC Phe	AAA Lys	TCA Ser	ATT Ile 520	Glu	GAC Asp	AAG Lys	CTA Leu	GGT Gly 525	ATC Ile	AAC Asn	TTT Phe	AAG Lys	ATT Ile 530	1701
GAG Glu	AGG Arg	TCC Ser	ATT Ile	GAT Asp 535	Asp	ATC Ile	AGG Arg	GGC Gly	AAG Lys 540	Leu	AGA Arg	CAG Gln	CTT Leu	GTC Val 545	CTC Leu	1749

	CCA Pro 550							1797
	GTT Val							1845
	GGG Gly							1893
	GCG Ala							1941
	ATC Ile							1989
	GGT Gly 630							2037
	GGC Gly							2085
	TTC Phe							2133
	GGC Gly							2181
	AAC Asn							2229
	ACT Thr 710							2277
	AGG Arg							2325
	AGC Ser							2373

GAG Glu 7 5 5	AAA Lys	CTC Leu	CAC His	AAG Lys	TCC Ser 760	AAG Lys	CCA Pro	GAC Asp	GAC Asp	CCC Pro 765	GAT Asp	GCA Ala	GAC Asp	TGG Trp	TTC Phe 770	2421
GAA Glu	AGA Arg	TCA Ser	GAA Glu	ACT Thr 775	CTG Leu	TCA Ser	GAC Asp	CTT Leu	CTG Leu 780	GAG Glu	AAA Lys	GCC Ala	GAC Asp	ATC Ile 785	GCC Ala	2469
AGC Ser	AAG Lys	GTC Val	GCC Ala 790	CAC His	TCA Ser	GCA Ala	CTC Leu	GTG Val 795	GAA Glu	ACA Thr	AGC Ser	GAC Asp	GCC Ala 800	CTT Leu	GAA Glu	2517
GCA Ala	GTT Val	CAG Gln 805	TCG Ser	ACT Thr	TCC Ser	GTG Val	TAC Tyr 810	ACC Thr	CCC Pro	AAG Lys	TAC Tyr	CCA Pro 815	GAA Glu	GTC Val	AAG Lys	2565
AAC Asn	CCA Pro 820	CAG Gln	ACC Thr	GCC Ala	TCC Ser	AAC Asn 825	CCC Pro	GTT Val	GTT Val	GGG Gly	CTC Leu 830	CAC His	CTG Leu	CCC Pro	GCC Ala	2613
AAG Lys 835	AGA Arg	GCC Ala	ACC Thr	GGT Gly	GTC Val 840	CAG Gln	GCC Ala	GCT Ala	CTT Leu	CTC Leu 845	GGA Gly	GCA Ala	GGA Gly	ACG Thr	AGC Ser 850	2661
AGA Arg	CCA Pro	ATG Met	GGG Gly	ATG Met 855	GAG Glu	GCC Ala	CCA Pro	ACA Thr	CGG Arg 860	TCC Ser	AAG Lys	AAC Asn	GCC Ala	GTG Val 865	AAA Lys	2709
ATG Met	GCC Ala	AAA Lys	CGG Arg 870	CGG	CAA Gln	CGC Arg	CAA Gln	AAG Lys 875	GAG Glu	AGC Ser	CGC Arg	TAA	CAGC	CAT		2755
GAT	GGA)	ACC 1	ACTC	AAGA	AG AG	GGAC	ACTA	A TC	CCAG	ACCC	CGT	ATCC(CCG (GCCT'	rcgcct	2815
GCG(GGGG	ccc (cc													2827

- (2) INFORMATION FOR SEQ ID NO:26:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 878 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Ser Asp Ile Phe Asn Ser Pro Gln Ala Arg Ser Thr Ile Ser Ala 5 . 1

Ala Phe Gly Ile Lys Pro Thr Ala Gly Gln Asp Val Glu Glu Leu Leu

			20					25					30		
Ile	Pro	Lys 35	Val	Trp	Val	Pro	Pro 40	Glu	Asp	Pro	Leu	Ala 45	Ser	Pro	Ser
Arg	Leu 50	Ala	Lys	Phe	Leu	Arg 55	Glu	Asn	Gly	Tyr	Lys 60	Val	Leu	Gln	Pro
Arg 65	Ser	Leu	Pro	Glu	Asn 70	Glu	Glu	Tyr	Glu	Thr 75	Asp	Gln	Ile	Leu	Pro 80
Asp	Leu	Ala	Trp	Met 85	Arg	Gln	Ile	Glu	Gly 90	Ala	Val	Leu	Lys	Pro 95	Thr
Leu	Ser	Leu	Pro 100	Ile	Gly	Asp	Gln	Glu 105	Tyr	Phe	Pro	Lys	Tyr 110	Tyr	Pro
Thr	His	Arg 115	Pro	Ser	Lys	Glu	Lys 120	Pro	Asn	Ala	Tyr	Pro 125	Pro	Asp	Ile
Ala	Leu 130	Leu	Lys	Gln	Met	Ile 135	Tyr	Leu	Phe	Leu	Gln 140	Val	Pro	Glu	Ala
Asn 145	Glu	Gly	Leu	Lys	Asp 150	Glu	Val	Thr	Leu	Leu 155	Thr	Gln	Asn	Ile	Arg 160
Asp	Lys	Ala	Tyr	Gly 165	Ser	Gly	Thr	Tyr	Met 170	Gly	Gln	Ala	Asn	Arg 175	Leu
Val	Ala	Met	Lys 180	Glu	Val	Ala	Thr	Gly 185	Arg	Asn	Pro	Asn	Lys 190	qaA	Pro
Leu	Lys	Leu 195	Gly	Tyr	Thr	Phe	Glu 200	Ser	Ile	Ala	Gln	Leu 205	Leu	Asp	Ile
Thr	Leu 210	Pro	Val	Gly	Pro	Pro 215	Gly	Glu	Asp	Asp	Lys 220	Pro	Trp	Val	Pro
Leu 225	Thr	Arg	Val	Pro	Ser 230	Arg	Met	Leu	Val	Leu 235	Thr	Gly	Asp	Val	Asp 240
Gly	Asp	Phe	Glu	Val 245	Glu	Asp	Tyr	Leu	Pro 250	Lys	Ile	Asn	Leu	Lys 255	Ser
Ser	Ser	Gly	Leu 260	Pro	Tyr	Val	Gly	Arg 265	Thr	Lys	Gly	Glu	Thr 270	Ile	Gly
Glu	Met	Ile 275	Ala	Ile	Ser	Asn	Gln 280	Phe	Leu	Arg	Glu	Leu 285	Ser	Thr	Leu
Leu	Lys	Gln	Gly	Ala	Gly	Thr	Lys	Gly	Ser	Asn	Lys	Lys	Lys	Leu	Leu

Ser Met Leu Ser Asp Tyr Trp Tyr Leu Ser Cys Gly Leu Leu Phe Pro Lys Ala Glu Arg Tyr Asp Lys Ser Thr Trp Leu Thr Lys Thr Arg Asn Ile Trp Ser Ala Pro Ser Pro Thr His Leu Met Ile Ser Met Ile Thr Trp Pro Val Met Ser Asn Ser Pro Asn Asn Val Leu Asn Ile Glu Gly Cys Pro Ser Leu Tyr Lys Phe Asn Pro Phe Arg Gly Gly Leu Asn Arg Ile Val Glu Trp Ile Leu Ala Pro Glu Glu Pro Lys Ala Leu Val Tyr Ala Asp Asn Ile Tyr Ile Val His Ser Asn Thr Trp Tyr Ser Ile Asp Leu Glu Lys Gly Glu Ala Asn Cys Thr Arg Gln His Met Gln Ala Ala Met Tyr Tyr Ile Leu Thr Arg Gly Trp Ser Asp Asn Gly Asp Pro Met Phe Asn Gln Thr Trp Ala Thr Phe Ala Met Asn Ile Ala Pro Ala Leu Val Val Asp Ser Ser Cys Leu Ile Met Asn Leu Gln Ile Lys Thr Tyr Gly Gln Gly Ser Gly Asn Ala Ala Thr Phe Ile Asn Asn His Leu Leu Ser Thr Leu Val Leu Asp Gln Trp Asn Leu Met Arg Gln Pro Arg Pro Asp Ser Glu Glu Phe Lys Ser Ile Glu Asp Lys Leu Gly Ile Asn Phe Lys Ile Glu Arg Ser Ile Asp Asp Ile Arg Gly Lys Leu Arg Gln Leu Val Leu Leu Ala Gln Pro Gly Tyr Leu Ser Gly Gly Val Glu Pro Glu Gln Ser Ser Pro Thr Val Glu Leu Asp Leu Leu Gly Trp Ser Ala Thr

				565					570					57 5	
Tyr	Ser	Lys	Asp 580	Leu	Gly	Ile	Tyr	Val 585	Pro	Val	Leu	Asp	Lys 590	Glu	Arg
Leu	Phe	Cys 595	Ser	Ala	Ala	Tyr	Pro 600	Lys	Gly	Val	Glu	Asn 605	Lys	Ser	Leu
Lys	Ser 610	Lys	Val	Gly	Ile	Glu 615	Gln	Ala	Tyr	Lys	Val 620	Val	Arg	Tyr	Glu
Ala 625	Leu	Arg	Leu	Val	Gly 630	Gly	Trp	Asn	Tyr	Pro 635	Leu	Leu	Asn	Lys	Ala 640
Cys	Lys	Asn	Asn	Ala 645	Gly	Ala	Ala	Arg	Arg 650	His	Leu	Glu	Ala	Lys 655	Gly
Phe	Pro	Leu	Asp 660	Glu	Phe	Leu	Ala	Glu 665	Trp	Ser	Glu	Leu	Ser 670	Glu	Phe
_	Glu	675					680		_			685			
	Leu 690					695					700				
705	Arg				710		-			715					720
-	Thr	-		725	-				730			_		735	
	Ala		740	_				745					750		
	Ala	755					760					765			_
-	Phe 770		_			775					780				_
785	Ala		-		790					795					800
Leu	Glu	Ala	Val	Gln 805	Ser	Thr	Ser	Val	Tyr 810	Thr	Pro	Lys	Tyr	Pro 815	Glu
Val	Lys	Asn	Pro 820	Gln	Thr	Ala	Ser	Asn 825	Pro	Val	Val	Gly	Leu 830	His	Leu
Pro	Ala	Lys	Arg	Ala	Thr	Gly	Val	Gln	Ala	Ala	Leu	Leu	Gly	Ala	Gly

37

845 840 835 Thr Ser Arg Pro Met Gly Met Glu Ala Pro Thr Arg Ser Lys Asn Ala 860 855 850 Val Lys Met Ala Lys Arg Arg Gln Arg Gln Lys Glu Ser, Arg 875 870 865 (2) INFORMATION FOR SEQ ID NO:27: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3261 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: circular (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 97..531 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27: GGATACGATC GGTCTGACCC CGGGGGAGTC ACCCGGGGAC AGGCCGTCAA GGCCTTGTTC 60 CAGGATGGGA CTCCTCCTTC TACAACGCTA TCATTG ATG GTT AGT AGA GAT CAG 114 Met Val Ser Arg Asp Gln 880 ACA AAC GAT CGC AGC GAT GAC AAA CCT GCA AGA TCA AAC CCA ACA GAT 162 Thr Asn Asp Arg Ser Asp Asp Lys Pro Ala Arg Ser Asn Pro Thr Asp 900 895 890 885 TGT TCC GTT CAT ACG GAG CCT TCT GAT GCC AAC AAC CGG ACC GGC GTC 210 Cys Ser Val His Thr Glu Pro Ser Asp Ala Asn Asn Arg Thr Gly Val 915 905 CAT TCC GGA CGA CAC CCT GGA GAA GCA CAC TCT CAG GTC AGA GAC CTC 258 His Ser Gly Arg His Pro Gly Glu Ala His Ser Gln Val Arg Asp Leu 925 920 GAC CTA CAA TTT GAC TGT GGG GGA CAC AGG GTC AGG GCT AAT TGT CTT 306 Asp Leu Gln Phe Asp Cys Gly Gly His Arg Val Arg Ala Asn Cys Leu 945 940 935 TTT CCC TGG ATT CCC TGG CTC AAT TGT GGG TGC TCA CTA CAC ACT GCA 354 Phe Pro Trp Ile Pro Trp Leu Asn Cys Gly Cys Ser Leu His Thr Ala

955

950

GGG CAA TGG GAA CTA CAA GTT CGA TCA GAT GCT CCT GAC TGC CCA GAA Gly Gln Trp Glu Leu Gln Val Arg Ser Asp Ala Pro Asp Cys Pro Glu 965 970 975 980	402
CCT ACC GGC CAG TTA CAA CTA CTG CAG GCT AGT GAG TCG GAG TCT CAC Pro Thr Gly Gln Leu Gln Leu Gln Ala Ser Glu Ser Glu Ser His 985	450
AGT GAG GTC AAG CAC ACT TCC TGG TGG CGT TTA TGC ACT AAA CGG CAC Ser Glu Val Lys His Thr Ser Trp Trp Arg Leu Cys Thr Lys Arg His 1000 1005 1010	498
CAT AAA CGC CGT GAC CTT CCA AGG AAG CCT GAG TGAACTGACA GATGTTAGCT His Lys Arg Arg Asp Leu Pro Arg Lys Pro Glu 1015 1020	551
ACAATGGGTT GATGTCTGCA ACAGCCAACA TCAACGACAA AATTGGGAAC GTCCTAGTAG	611
GGGAAGGGT CACCGTCCTC AGCTTACCCA CATCATATGA TCTTGGGTAT GTGAGGCTTG	671
GTGACCCCAT TCCCGCAATA GGGCTTGACC CAAAAATGGT AGCCACATGT GACAGCAGTG	731
ACAGGCCCAG AGTCTACACC ATAACTGCAG CCGATGATTA CCAATTCTCA TCACAGTACC	791
AACCAGGTGG GGTAACAATC ACACTGTTCT CAGCCAACAT TGATGCCATC ACAAGCCTCA	851
GCGTTGGGGG AGAGCTCGTG TTTCAAACAA GCGTCCACGG CCTTGTACTG GGCGCCACCA	911
TCTACCTCAT AGGCTTTGAT GGGACAACGG TAATCACCAG GGCTGTGGCC GCAAACAATG	971
GGCTGACGAC CGGCACCGAC AACCTTATGC CATTCAATCT TGTGATTCCA ACAAACGAGA	1031
TAACCCAGCC AATCACATCC ATCAAACTGG AGATAGTGAC CTCCAAAAGT GGTGGTCAGG	1091
CAGGGGATCA GATGTCATGG TCGGCAAGAG GGAGCCTAGC AGTGACGATC CATGGTGGCA	1151
ACTATCCAGG GGCCCTCCGT CCCGTCACGC TAGTGGCCTA CGAAAGAGTG GCAACAGGAT	1211
CCGTCGTTAC GGTCGCTGGG GTGAGCAACT TCGAGCTGAT CCCAAATCCT GAACTAGCAA	1271
AGAACCTGGT TACAGAATAC GGCCGATTTG ACCCAGGAGC CATGAACTAC ACAAAATTGA	1331
FACTGAGTGA GAGGGACCGT CTTGGCATCA AGACCGTCTG GCCAACAAGG GAGTACACTG	1391
ACTTTCGTGA ATACTTCATG GAGGTGGCCG ACCTCAACTC TCCCCTGAAG ATTGCAGGAG	1451
CATTCGGCTT CAAAGACATA ATCCGGGCCA TAAGGAGGAT AGCTGTGCCG GTGGTCTCCA	
CATTGTTCCC ACCTGCCGCT CCCCTAGCCC ATGCAATTGG GGAAGGTGTA GACTACCTGC	
rgggcgatga ggcacaggct gcttcaggaa ctgctcgagc cgcgtcagga aaagcaagag	

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CTGCCTCAGG	CCGCATAAGG	CAGCTGACTC	TCGCCGCCGA	CAAGGGGTAC	GAGGTAGTCG	1691
CGAATCTATT	CCAGGTGCCC	CAGAATCCCG	TAGTCGACGG	GATTCTTGCT	TCACCTGGGG	1751
TACTCCGCGG	TGCACACAAC	CTCGACTGCG	TGTTAAGAGA	GGGTGCCACG	CTATTCCCTG	1811
rggttattac	GACAGTGGAA	GACGCCATGA	CACCCAAAGC	ATTGAACAGC	AAAATGTTTG	1871
CTGTCATTGA	AGGCGTGCGA	GAAGACCTCC	AACCTCCATC	TCAAAGAGGA	TCCTTCATAC	1931
GAACTCTCTC	TGGACACAGA	GTCTATGGAT	ATGCTCCAGA	TGGGGTACTT	CCACTGGAGA	1991
CTGGGAGAGA	CTACACCGTT	GTCCCAATAG	ATGATGTCTG	GGACGACAGC	ATTATGCTGT	2051
CCAAAGATCC	CATACCTCCT	ATTGTGGGAA	ACAGTGGAAA	TCTAGCCATA	GCTTACATGG	2111
ATGTGTTTCG	ACCCAAAGTC	CCAATCCATG	TGGCTATGAC	GGGAGCCCTC	AATGCTTGTG	2171
GCGAGATTGA	GAAAGTAAGC	TTTAGAAGCA	CCAAGCTCGC	CACTGCACAC	CGACTTGGCC	2231
TTAGGTTGGC	TGGTCCCGGA	GCATTCGATG	TAAACACCGG	GCCCAACTGG	GCAACGTTCA	2291
TCAAACGTTT	CCCTCACAAT	CCACGCGACT	GGGACAGGCT	CCCCTACCTC	AACCTACCAT	2351
ACCTTCCACC	CAATGCAGGA	CGCCAGTACC	ACCTTGCCAT	GGCTGCATCA	GAGTTCAAAG	2411
AGACCCCCGA	ACTCGAGAGT	GCCGTCAGAG	CAATGGAAGC	AGCAGCCAAC	GTGGACCCAC	2471
TATTCCAATC	TGCACTCAGT	GTGTTCATGT	GGCTGGAAGA	GAATGGGATT	GTGACTGACA	2531
TGGCCAACTT	CGCACTCAGC	GACCCGAACG	CCCATCGGAT	GCGAAATTTT	CTTGCAAACG	2591
CACCACAAGC	AGGCAGCAAG	TCGCAAAGGG	CCAAGTACGG	GACAGCAGGC	TACGGAGTGG	2651
AGGCTCGGGG	CCCCACACCA	GAGGAAGCAC	AGAGGGAAAA	AGACACACGG	ATCTCAAAGA	2711
AGATGGAGAC	CATGGGCATC	TACTTTGCAA	CACCAGAATG	GGTAGCACTC	AATGGGCACC	2771
GAGGGCCAAG	CCCCGGCCAG	CTAAAGTACT	GGCAGAACAC	ACGAGAAATA	CCGGACCCAA	2831
ACGAGGACTA	TCTAGACTAC	GTGCATGCAG	AGAAGAGCCG	GTTGGCATCA	GAAGAACAAA	2891
TCCTAAGGG	AGCTACGTCG	ATCTACGGGG	CTCCAGGACA	GGCAGAGCCA	CCCCAAGCTT	2951
TCATAGACGA	AGTTGCCAAA	GTCTATGAAA	TCAACCATGG	ACGTGGCCCA	AACCAAGAAC	3011
AGATGAAAGA	A TCTGCTCTTG	; ACTGCGATGG	AGATGAAGCA	A TCGCAATCCC	AGGCGGGCTC	3071
TACCAAAGC	CAAGCCAAAA	CCCAATGCTC	CAACACAGAG	ACCCCCTGG1	CGGCTGGGCC	3131
GCTGGATCAG	GACCGTCTCT	GATGAGGACC	TTGAGTGAGG	CTCCTGGGAG	TCTCCCGACA	3191

40

CCACCCGCGC AGGTGTGGAC ACCAATTCGG CCTTACAACA TCCCAAATTG GATCCGTTCG 3251
CGGGTCCCCT 3261

- (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 145 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Ala 1 5 10 15

Arg Ser Asn Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala
20 25 30

Asn Asn Arg Thr Gly Val His Ser Gly Arg His Pro Gly Glu Ala His
35 40 45

Ser Gln Val Arg Asp Leu Asp Leu Gln Phe Asp Cys Gly Gly His Arg 50 55 60

Val Arg Ala Asn Cys Leu Phe Pro Trp Ile Pro Trp Leu Asn Cys Gly 65 70 75 80

Cys Ser Leu His Thr Ala Gly Gln Trp Glu Leu Gln Val Arg Ser Asp
85 90 95

Ala Pro Asp Cys Pro Glu Pro Thr Gly Gln Leu Gln Leu Gln Ala
100 105 110

Ser Glu Ser Glu Ser His Ser Glu Val Lys His Thr Ser Trp Trp Arg

Leu Cys Thr Lys Arg His His Lys Arg Arg Asp Leu Pro Arg Lys Pro 130 135 140

Glu

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:

41

(A)	LENGTH: 3261 base pairs
(B)	TYPE: nucleic acid
(C)	STRANDEDNESS: single
(D)	TOPOLOGY: circular

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 131..3166

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

GGATACGATC GGTCTGACCC CGGGGGAGTC ACCCGGGGAC AGGCCGTCAA GGCCTTGTTC	60
CAGGATGGGA CTCCTCCTTC TACAACGCTA TCATTGATGG TTAGTAGAGA TCAGACAAAC	120
GATCGCAGCG ATG ACA AAC CTG CAA GAT CAA ACC CAA CAG ATT GTT CCG Met Thr Asn Leu Gln Asp Gln Thr Gln Gln Ile Val Pro 150 155	169
TTC ATA CGG AGC CTT CTG ATG CCA ACA ACC GGA CCG GCG TCC ATT CCG Phe Ile Arg Ser Leu Leu Met Pro Thr Thr Gly Pro Ala Ser Ile Pro 160 165 170	217
GAC GAC ACC CTG GAG AAG CAC ACT CTC AGG TCA GAG ACC TCG ACC TAC Asp Asp Thr Leu Glu Lys His Thr Leu Arg Ser Glu Thr Ser Thr Tyr 175	265
AAT TTG ACT GTG GGG GAC ACA GGG TCA GGG CTA ATT GTC TTT TTC CCT Asn Leu Thr Val Gly Asp Thr Gly Ser Gly Leu Ile Val Phe Pro 195 200 205	313
GGA TTC CCT GGC TCA ATT GTG GGT GCT CAC TAC ACA CTG CAG GGC AAT Gly Phe Pro Gly Ser Ile Val Gly Ala His Tyr Thr Leu Gln Gly Asn 210 215 220	361
GGG AAC TAC AAG TTC GAT CAG ATG CTC CTG ACT GCC CAG AAC CTA CCG Gly Asn Tyr Lys Phe Asp Gln Met Leu Leu Thr Ala Gln Asn Leu Pro 225 230 235	409
GCC AGT TAC AAC TAC TGC AGG CTA GTG AGT CGG AGT CTC ACA GTG AGG Ala Ser Tyr Asn Tyr Cys Arg Leu Val Ser Arg Ser Leu Thr Val Arg 240 245 250	457
TCA AGC ACA CTT CCT GGT GGC GTT TAT GCA CTA AAC GGC ACC ATA AAC Ser Ser Thr Leu Pro Gly Gly Val Tyr Ala Leu Asn Gly Thr Ile Asn 265 270	505
GCC GTG ACC TTC CAA GGA AGC CTG AGT GAA CTG ACA GAT GTT AGC TAC	553

Ala	Val	Thr	Phe	Gln 275	Gly	Ser	Leu	Ser	Glu 280	Leu	Thr	Asp	Val	Ser 285	Tyr	
														GGG Gly		601
														TCA Ser		649
														GGG Gly		697
														AGA Arg		745
														TAC Tyr 365		793
														GCC Ala		841
														GTC Val		889
														GGG Gly		937
														ACC Thr		985
														GAG Glu 445		1033
														AAA Lys		1081
														AGC Ser		1129

GCA Ala	GTG Val 480	ACG Thr	ATC Ile	CAT His	GGT Gly	GGC Gly 485	AAC Asn	TAT Tyr	CCA Pro	GGG Gly	GCC Ala 490	CTC Leu	CGT Arg	CCC Pro	GTC Val	1177
ACG Thr 495	CTA Leu	GTG Val	GCC Ala	TAC Tyr	GAA Glu 500	AGA Arg	GTG Val	GCA Ala	ACA Thr	GGA Gly 505	TCC Ser	GTC Val	GTT Val	ACG Thr	GTC Val 510	1225 ·
GCT Ala	GGG Gly	GTG Val	AGC Ser	AAC Asn 515	TTC Phe	GAG Glu	CTG Leu	ATC Ile	CCA Pro 520	AAT Asn	CCT Pro	GAA Glu	CTA Leu	GCA Ala 525	AAG Lys	1273
AAC Asn	CTG Leu	GTT Val	ACA Thr 530	GAA Glu	TAC Tyr	GGC	CGA Arg	TTT Phe 535	GAC Asp	CCA Pro	GGA Gly	GCC Ala	ATG Met 540	AAC Asn	TAC Tyr	1321
ACA Thr	AAA Lys	TTG Leu 545	ATA Ile	CTG Leu	AGT Ser	GAG Glu	AGG Arg 550	GAC Asp	CGT Arg	CTT Leu	GGC Gly	ATC Ile 555	AAG Lys	ACC Thr	GTC Val	1369
TGG Trp	CCA Pro 560	ACA Thr	AGG Arg	GAG Glu	TAC Tyr	ACT Thr 565	GAC Asp	TTT Phe	CGT Arg	GAA Glu	TAC Tyr 570	TTC Phe	ATG Met	GAG Glu	GTG Val	1417
GCC Ala 575	GAC Asp	CTC Leu	AAC Asn	TCT Ser	CCC Pro 580	CTG Leu	AAG Lys	ATT Ile	GCA Ala	GGA Gly 585	GCA Ala	TTC Phe	GGC Gly	TTC Phe	AAA Lys 590	1465
GAC Asp	ATA Ile	ATC Ile	CGG Ar g	GCC Ala 595	ATA Ile	AGG Arg	AGG Arg	ATA Ile	GCT Ala 600	GTG Val	CCG Pro	GTG Val	GTC Val	TCC Ser 605	ACA Thr	1513
TTG Leu	TTC Phe	CCA Pro	CCT Pro 610	GCC Ala	GCT Ala	CCC Pro	CTA Leu	GCC Ala 615	CAT His	GCA Ala	ATT Ile	GGG Gly	GAA Glu 620	GGT Gly	GTA Val	1561
GAC Asp	TAC Tyr	CTG Leu 625	CTG Leu	GGC Gly	GAT Asp	GAG Glu	GCA Ala 630	CAG Gln	GCT Ala	GCT Ala	TCA Ser	GGA Gly 635	ACT Thr	GCT Ala	CGA Arg	1609
GCC Ala	GCG Ala 640	TCA Ser	GGA Gly	AAA Lys	GCA Ala	AGA Arg 645	GCT Ala	GCC Ala	TCA Ser	GGC Gly	CGC Arg 650	ATA Ile	AGG Arg	CAG Gln	CTG Leu	1657
ACT Thr 655	Leu	GCC Ala	GCC Ala	GAC Asp	AAG Lys 660	GGG Gly	TAC Tyr	GAG Glu	GTA Val	GTC Val 665	GCG Ala	AAT Asn	CTA Leu	TTC Phe	CAG Gln 670	1705
GTG Val	CCC Pro	CAG Gln	AAT Asn	CCC Pro 675	Val	GTC Val	GAC Asp	GGG Gly	ATT Ile 680	Leu	GCT Ala	TCA Ser	CCT Pro	GGG Gly 685	GTA Val	1753

PCT/US97/12955

					AAC Asn											1801
CTA Leu	TTC Phe	CCT Pro 705	GTG Val	GTT Val	ATT Ile	ACG Thr	ACA Thr 710	GTG Val	GAA Glu	GAC Asp	GCC Ala	ATG Met 715	ACA Thr	CCC Pro	AAA Lys	1849
GCA Ala	TTG Leu 720	AAC Asn	AGC Ser	AAA Lys	ATG Met	TTT Phe 725	GCT Ala	GTC Val	ATT Ile	GAA Glu	GGC Gly 730	GTG Val	CGA Arg	GAA Glu	GAC Asp	1897
					CAA Gln 740											1945
					TAT Tyr											1993
					GTT Val											2041
					GAT Asp											2089
					TAC Tyr											2137
					GGA Gly 820											2185
					ACC Thr											2233
					GGA Gly											2281
					CGT Arg											2329
					CTA Leu											2377

TAC Tyr 895	CAC His	CTT Leu	GCC Ala	ATG Met	GCT Ala 900	GCA Ala	TCA Ser	GAG Glu	TTC Phe	AAA Lys 905	GAG Glu	ACC Thr	CCC Pro	GAA Glu	CTC Leu 910	2425
GAG Glu	AGT Ser	GCC Ala	GTC Val	AGA Arg 915	GCA Ala	ATG Met	GAA Glu	GCA Ala	GCA Ala 920	GCC Ala	AAC Asn	GTG Val	GAC Asp	CCA Pro 925	CTA Leu	2473
TTC Phe	CAA Gln	TCT Ser	GCA Ala 930	CTC Leu	AGT Ser	GTG Val	TTC Phe	ATG Met 935	TGG Trp	CTG Leu	GAA Glu	GAG Glu	AAT Asn 940	GGG Gly	ATT Ile	2521
Val	ACT Thr	Asp 945	Met	Ala	Asn	Phe	Ala 950	Leu	Ser	Asp	Pro	Asn 955	Ala	His	Arg	2569
ATG Met	CGA Arg 960	AAT Asn	TTT Phe	CTT Leu	GCA Ala	AAC Asn 965	GCA Ala	CCA Pro	CAA Gln	GCA Ala	GGC Gly 970	AGC Ser	AAG Lys	TCG Ser	CAA Gln	2617
Arg 975	GCC Ala	Lys	Tyr	Gly	Thr 980	Ala	Gly	Tyr	Gly	Val 985	Glu	Ala	Arg	GIA	990	2665
ACA Thr	CCA Pro	GAG Glu	GAA Glu	GCA Ala 995	CAG Gln	AGG Arg	GAA Glu	AAA Lys	GAC Asp 100	Thr	CGG Arg	ATC Ile	TCA Ser	AAG Lys 100	Lys	2713
ATG Met	GAG Glu	ACC Thr	ATG Met 101	Gly	ATC Ile	TAC Tyr	TTT Phe	GCA Ala 101	Thr	CCA Pro	GAA Glu	TGG Trp	GTA Val 102	Ala	CTC Leu	2761
AAT Asn	GGG Gly	CAC His 102	Arg	GGG	CCA Pro	AGC Ser	CCC Pro 103	Gly	CAG Gln	CTA Leu	AAG Lys	TAC Tyr 103	Trp	CAG Gln	AAC Asn	2809
ACA Thr	CGA Arg 104	Glu	ATA Ile	CCG Pro	GAC Asp	CCA Pro 104	Asn	GAG Glu	GAC Asp	TAT Tyr	CTA Leu 105	Asp	TAC	GTG Val	CAT His	2857
GCA Ala 105	Glu	AAG Lys	AGC Ser	CGG Arg	TTG Leu 106	Ala	TCA Ser	GAA Glu	GAA Glu	CAA Gln 106	Ile	CTA Leu	AGG Arg	GCA Ala	GCT Ala 1070	2905
ACG Thr	TCG Ser	ATC	TAC Tyr	GGG Gly 107	Ala	CCA Pro	GGA Gly	CAG Gln	GCA Ala 108	Glu	CCA Pro	CCC Pro	CAA Gln	GCT Ala 108	TTC Phe 5	2953
ATA	GAC Asp	GAA	GTT Val	Ala	: AAA Lys	GTC Val	TAT	GAA Glu 109	ılle	AAC Asr	CAT His	GGA Gly	CGT Arg	f Gly	CCA Pro	3001

AA Ası	CAA 1 Gln	GAA Glu 110	ı Gln	ATG Met	AAA Lys	GAT Asp	CTG Leu 111	Lev	TTG Leu	ACT Thr	GCG Ala	ATO Met	: Glu	ATO Met	AAG Lys	3049
CAT	CGC Arg	Asr	CCC Pro	AGG Arg	CGG Arg	GCT Ala 112	Leu	CCA Pro	AAG Lys	CCC Pro	AAG Lys 113	Pro	AAA Lys	CCC	AAT Asn	3097
GCT Ala 113	Pro	ACA Thr	CAG Gln	AGA Arg	CCC Pro 114	Pro	GGT Gly	CGG Arg	CTG Leu	GGC Gly 114	Arg	TGG Trp	ATC Ile	AGG Arg	ACC Thr 1150	3145
			GAG Glu		Leu		TGA	GGCT	CCT	GGGA	GTCT	CC C	GACA	CCAC	C	3196
CGC	GCAG	GTG	TGGA	CACC	AA T	rcgg	CCTT	A CA	ACAT	CCCA	AAT'	TGGA	TCC (GTTC	GCGGGT	3256
CCC	CT															3261
(2)	(i)	(i)	(B)	ENCE) LEN) TYI) TOI	CHAI NGTH: PE: & POLOG	RACTE : 101 amino 3Y: 1	ERIST 12 ar 2 ac: 1 inea	rics mino id ar	acio		30 :					
Met 1	Thr	Asn	Leu	Gln 5	Asp	Gln	Thr	Gln	Gln 10	Ile	Val	Pro	Phe	Ile 15	Arg	
Ser	Leu	Leu	Met 20	Pro	Thr	Thr	Gly	Pro 25	Ala	Ser	Ile	Pro	Asp 30	Asp	Thr	
Leu	Glu	Lys 35	His	Thr	Leu	Arg	Ser 40	Glu	Thr	Ser	Thr	Tyr 45	Asn	Leu	Thr	
Val	Gly 50	Asp	Thr	Gly	Ser	Gly 55	Leu	Ile	Val	Phe	Phe 60	Pro	Gly	Phe	Pro	
Gly 65	Ser	Ile	Val	Gly	Ala 70	His	Tyr	Thr	Leu	Gln 75	Gly	Asn	Gly	Asn	Tyr 80	
Lys	Phe	Asp	Gln	Met 85	Leu	Leu	Thr	Ala	Gln 90	Asn	Leu	Pro	Ala	Ser 95	Tyr	
Asn	Tyr	Cys	Arg 100	Leu	Val	Ser	Arg	Ser 105	Leu	Thr	Val	Arg	Ser 110	Ser	Thr	

Leu	Pro	Gly 115	Gly	Val	Tyr	Ala	Leu 120	Asn	Gly	Thr	Ile	Asn 125	Ala	Val	Thr
Phe	Gln 130	Gly	Ser	Leu	Ser	Glu 135	Leu	Thr	Asp	Val	Ser 140	Tyr	Asn	Gly	Leu
Met 145	Ser	Ala	Thr	Ala	Asn 150	Ile	Asn	Asp	Lys	Ile 155	Gly	Asn	Val	Leu	Val 160
Gly	Glu	Gly	Val	Thr 165	Val	Leu	Ser	Leu	Pro 170	Thr	Ser	Tyr	Asp	Leu 175	Gly
-			Leu 180					185					190		
		195	Thr				200					205			
	210		Asp			215					220				
225			Thr		230					235					240
			Gly	245					250					255	
			Thr 260					265					270		
		275	Val				280					285			
Leu	Met 290	Pro	Phe	Asn	Leu	Val 295	Ile	Pro	Thr	Asn	Glu 300	Ile	Thr	Gln	Pro
305			Ile		310					315					320
			Gln	325					330					335	
Ile	His	Gly	Gly 340	Asn	Tyr	Pro	Gly	Ala 345	Leu	Arg	Pro	Val	Thr 350	Leu	Val
Ala	Tyr	Glu 355	Arg	Val	Ala	Thr	Gly 360	Ser	Val	Val	Thr	Val 365	Ala	Gly	Val
Ser	Asn 370		Glu	Leu	Ile	Pro 375	Asn	Pro	Glu	Leu	Ala 380	Lys	Asn	Leu	Val
mb	~1	Т	. C1v	ስተማ	Dha	Agn	Pro	Glv	Ala	Met	Asn	Tyr	Thr	Lvs	Let

385					390					395					400
Ile	Leu	Ser	Glu	Arg 405	Asp	Arg	Leu	Gly	Ile 410	Lys	Thr	Val	Trp	Pro 415	Thr
Arg	Glu	Tyr	Thr 420	Asp	Phe	Arg	Glu	Tyr 425	Phe	Met	Glu	Val	Ala 430	Asp	Leu
A sn	Ser	Pro 435	Leu	Lys	Ile	Ala	Gly 440	Ala	Phe	Gly	Phe	Lys 445	Asp	Ile	Ile
Arg	Ala 450	Ile	Arg	Arg	Ile	Ala 455	Val	Pro	Val	Val	Ser 460	Thr	Leu	Phe	Pro
Pro 465	Ala	Ala	Pro	Leu	Ala 470	His	Ala	Ile	Gly	Glu 475	Gly	Val	Asp	Tyr	Leu 480
Leu	Gly	Asp	Glu	Ala 485	Gln	Ala	Ala	Ser	Gly 490	Thr	Ala	Arg	Ala	Ala 495	Ser
Gly	Lys	Ala	Arg 500	Ala	Ala	Ser	Gly	Ar g 505	Ile	Arg	Gln	Leu	Thr 510	Leu	Ala
Ala	Asp	Lys 515	Gly	Tyr	Glu	Val	Val 520	Ala	Asn	Leu	Phe	Gln 525	Val	Pro	Gln
Asn	Pro 530	Val	Val	Asp	Gly	Ile 535	Leu	Ala	Ser	Pro	Gly 540	Val	Leu	Arg	Gly
Ala 545	His	Asn	Leu	Asp	Cys 550	Val	Leu	Arg	Glu	Gly 555	Ala	Thr	Leu	Phe	Pro 560
Val	Val	Ile	Thr	Thr 565	Val	Glu	Asp	Ala	Met 570	Thr	Pro	Lys	Ala	Leu 575	Asn
Ser	Lys	Met	Phe 580	Ala	Val	Ile	Glu	Gly 585	Val	Arg	Glu	Asp	Leu 590	Gln	Pro
Pro	Ser	Gln 595	Arg	Gly	Ser	Phe	Ile 600	Arg	Thr	Leu	Ser	Gly 605	His	Arg	Val
Tyr	Gly 610	Tyr	Ala	Pro	Asp	Gly 615	Val	Leu	Pro	Leu	Glu 620	Thr	Gly	Arg	Asp
Tyr 625	Thr	Val	Val	Pro	Ile 630	Asp	Asp	Val	Trp	Asp 635	Asp	Ser	Ile	Met	Leu 640
Ser	Lys	Asp	Pro	Ile 645	Pro	Pro	Ile	Val	Gly 650	Asn	Ser	Gly	Asn	Leu 655	Ala
Ile	Ala	Tyr	Met 660	Asp	Val	Phe	Arg	Pro 665	Lys	Val	Pro	Ile	His 670	Val	Ala

PCT/US97/12955 WO 98/09646

- Met Thr Gly Ala Leu Asn Ala Cys Gly Glu Ile Glu Lys Val Ser Phe
- Arg Ser Thr Lys Leu Ala Thr Ala His Arg Leu Gly Leu Arg Leu Ala
- Gly Pro Gly Ala Phe Asp Val Asn Thr Gly Pro Asn Trp Ala Thr Phe
- Ile Lys Arg Phe Pro His Asn Pro Arg Asp Trp Asp Arg Leu Pro Tyr
- Leu Asn Leu Pro Tyr Leu Pro Pro Asn Ala Gly Arg Gln Tyr His Leu
- Ala Met Ala Ala Ser Glu Phe Lys Glu Thr Pro Glu Leu Glu Ser Ala
- Val Arg Ala Met Glu Ala Ala Ala Asn Val Asp Pro Leu Phe Gln Ser
- Ala Leu Ser Val Phe Met Trp Leu Glu Glu Asn Gly Ile Val Thr Asp
- Met Ala Asn Phe Ala Leu Ser Asp Pro Asn Ala His Arg Met Arg Asn
- Phe Leu Ala Asn Ala Pro Gln Ala Gly Ser Lys Ser Gln Arg Ala Lys
- Tyr Gly Thr Ala Gly Tyr Gly Val Glu Ala Arg Gly Pro Thr Pro Glu
- Glu Ala Gln Arg Glu Lys Asp Thr Arg Ile Ser Lys Lys Met Glu Thr
- Met Gly Ile Tyr Phe Ala Thr Pro Glu Trp Val Ala Leu Asn Gly His
- Arg Gly Pro Ser Pro Gly Gln Leu Lys Tyr Trp Gln Asn Thr Arg Glu
- Ile Pro Asp Pro Asn Glu Asp Tyr Leu Asp Tyr Val His Ala Glu Lys
- Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser Ile
- Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp Glu
- Val Ala Lys Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln Glu

945					950					955					960	
Gln	Met	Lys	Asp	Leu 965	Leu	Leu	Thr	Ala	Met 970	Glu	Met	Lys	His	Arg 975	Asn	
Pro	Arg	Arg	Ala 980	Leu	Pro	Lys	Pro	Lys 985	Pro	Lys	Pro	Asn	Ala 990	Pro	Thr	
Gln	Arg	Pro 995	Pro	Gly	Arg	Leu	Gly 1000		Trp	Ile	Arg	Thr 1009		Ser	Asp	,
Glu	Asp 1010		Glu													
(2)	INFO	RMAI	rion	FOR	SEQ	ID N	IO:31	L:								
		(<i>F</i> (E (C	QUENC A) LE B) TY C) SI D) TO	ENGTH PE: TRANI POLC	H: 32 nucl EDNE OGY:	eic Ess: circ	ase acid sing cular	pair l gle	ទ							
	(ii)	MOI	ECUI	E TY	PE:	CDNA	\									
	(ix)	(A	TURE NA LC	ME/K			531									
	(xi)	SEC	UENC	E DE	SCRI	PTIC	N: S	SEQ I	D NC	31:	:					
GGAI	ACGA	TC G	GTCI	GACC	C CG	GGGG	AGTO	ACC	CGGG	GAC	AGGC	CATO	AC I	GCCT	TGTTC	60
CTGG	TTGG	AA C	TCCT	CTTT	'C TG	CTGI	'ACTA	A TCG	STTG				Arg	GAT Asp		114
		Asp	CGC Arg				Lys					His				162
	Ser		CAT His			Pro					Asp					210
			CGA Arg		Pro					Thr					Leu	258
GAC	TTA	CAA	CTT	GAC	TGT	AGG	GGA	TAC	AGG	GTC	AGG	ACT	AAT	TGT	СТТ	306

qaA	Leu	Gln	Leu 1070		Сув	Arg	Gly	Tyr 1075		Val	Arg	Thr	Asn 108	Cys 0	Leu	
TTT Phe	CCC Pro	TGG Trp 1085	Ile	CCC Pro	TGG Trp	TTC Phe	AGT Ser 1090	Сув	AGG Arg	TGC Cys	TCA Ser	CTA Leu 1095	His	ACT Thr	GCA Ala	354
GAG Glu	CAG Gln 1100	Trp	GAA Glu	CTA Leu	CCA Pro	ATT Ile 1105	Arg	CCA Pro	GAT Asp	GCT Ala	CCT Pro 1110	Asp	AGC Ser	GCA Ala	GAA Glu	402
CCT Pro 1115	Ala	TGC Cys	CAG Gln	CTA Leu	CAA Gln 1120	Leu	CTG Leu	CAG Gln	GCT Ala	AGT Ser 1125	Glu	CAG Gln	GAG Glu	TCT Ser	AAC Asn 1130	450
CGT Arg	ACG Thr	GTC Val	AAG Lys	CAC His 1135	Thr	CCC Pro	TGG Trp	TGG Trp	CGT Arg 1140	Leu	TGC Cys	ACT Thr	AAA Lys	CGG Arg 114	Asn	498
CAT His	AAA Lys	CGC Arg	AGT Ser	Asp	CTT Leu	CCA Pro	CGG Arg	AAG Lys 1159	Pro	GAG Glu	TGAC	ettg <i>i</i>	ACT	GACT	ACAGCT	551
ACAZ	ACGG(GCT (GATG	rcago	CC AC	CTGC	BAAC	A TC	AACG/	ACAA	GAT	:GGG2	AAC	GTTC	ragttg	611
GAG	AAGG(GT (SACTO	GTTC	rc A	STCT	ACCG!	A CT	rcat;	ATGA	CCT	ragt:	TAT	GTGA	GACTCG	671
GTG	ACCC	CAT (cccc	GCAG	CA GO	SACT	CGAC	C CG2	AAGT:	rgat	GGC	CACG'	rgc	GACA	GTAGTG	731
ACAG	BACC	CAG A	AGTC1	raca(CC A	TAAC	AGCT	G CAC	GATG	AATA	CCA	ATTC:	rcg	TCAC	AACTCA	791
TCC	CGAG!	rgg (CGTG	AAGA(CC A	CACTO	GTTC	r cc	GCCA	ACAT	CGA:	rgct	CTC	ACCA	GCTTCA	851
GCG	rtgg:	rgg :	rgago	CTTG	rc T	rcag	CCAA	G TA	ACGA:	rcca	AAG	CATT	GAA	GTGG.	ACGTCA	911
CCA'	TTCA(CTT (CATT	GGGT"	rt G	ACGG	GACA	G AC	GTAG	CAGT	CAA	GCA(GTT	GCAA	CAGACT	971
TTG	GGCT	GAC 2	AACT	GGGA	CA A	ACAA	CCTT	G TG	CCAT	rcaa	CCT	GTG	GTC	CCAA	CAAATG	1031
AGA'	TCAC	CCA (GCCC	ATCA	CT T	CCAT	GAAA	C TA	GAGG'	TTGT	GAC	CTAC	AAG	ATTG	GCGGCA	1091
CCG	CTGG'	TGA	CCCA	ATAT	CA T	GGAC	AGTG.	A GT	GGTA	CACT	AGC'	rgtg.	ACG	GTGC	ACGGAG	1151
GCA	ACTA	CCC	TGGG	GCTC	TC C	GTCC'	TGTC.	A CC	CTGG'	TGGC	CTA'	rgaa:	CGA	GTGG	CTGCAG	1211
															AGCTTG	1271
															CCAAAC	1331
															AGTACA	
															TTGCAG	

GAGCATTTGG	CTTTAAGGAC	ATAATCCGAG	CCATTCGGAA	GATTGCGGTG	CCAGTGGTAT	1511
CCACACTCTT	CCCTCCAGCT	GCACCCCTAG	CACATGCAAT	CGGAGAAGGT	GTAGACTACC	1571
TCCTGGGCGA	CGAGGCCCAA	GCAGCCTCAG	GGACAGCTCG	AGCCGCGTCA	GGAAAAGCTA	1631
GAGCTGCCTC	AGGACGAATA	AGGCAGCTAA	CTCTCGCAGC	TGACAAGGGG	TGCGAGGTAG	1691
TCGCCAACAT	GTTCCAGGTG	CCCCAGAATC	CCATTGTTGA	TGGCATTCTG	GCATCCCCAG	1751
GAATCCTGCG	TGGCGCACAC	AACCTCGACT	GCGTGCTATG	GGAGGGAGCC	ACTCTTTTCC	1811
CTGTTGTCAT	TACGACACTC	GAGGATGAGC	TGACCCCCAA	GGCACTGAAC	AGCAAAATGT	1871
TTGCTGTCAT	TGAAGGTGTG	CGAGAGGACC	TCCAGCCTCC	ATCCCAACGG	GGATCCTTCA	1931
TTCGAACTCT	CTCTGGCCAT	AGAGTCTATG	GCTATGCCCC	AGACGGAGTA	CTGCCTCTGG	1991
AGACCGGGAG	AGACTACACC	GTTGTCCCAA	TTGATGATGT	GTGGGACGAT	AGCATAATGC	2051
TGTCGCAGGA	CCCCATACCT	CCAATCATAG	GGAACAGCGG	CAACCTAGCC	ATAGCATACA	2111
TGGATGTCTT	CAGGCCCAAG	GTCCCCATCC	ACGTGGCTAT	GACAGGGGCC	CTCAATGCCC	2171
GCGGTGAGAT	CGAGAGTGTT	ACGTTCCGCA	GCACCAAACT	CGCCACAGCC	CACCGACTTG	2231
GCATGAAGTT	AGCTGGTCCT	GGAGCCTATG	ACATTAATAC	AGGACCTAAC	TGGGCAACGT	2291
TCGTCAAACG	TTTCCCTCAC	AATCCCCGAG	ACTGGGACAG	GTTGCCCTAC	CTCAACCTTC	2351
CTTATCTCCC	ACCAACAGCA	GGACGTCAGT	TCCATCTAGC	CCTGGCTGCC	TCCGAGTTCA	2411
AAGAGACCCC	AGAACTCGAA	GACGCTGTGC	GCGCAATGGA	TGCCGCTGCA	AATGCCGACC	2471
CATTGTTCCG	CTCAGCTCTC	CAGGTCTTCA	TGTGGTTGGA	AGAAAACGGG	ATTGTGACCG	2531
ACATGGCTAA	CTTCGCCCTC	AGCGACCCAA	ACGCGCATAG	GATGAAAAAC	TTCCTAGCAA	2591
ACGCACCCCA	GGCTGGAAGC	AAGTCGCAGA	GGGCCAAGTA	TGGCACGGCA	GGCTACGGAG	2651
TGGAGGCTCG	AGGCCCCACA	CCAGAAGAGG	CACAGAGGGA	AAAAGACACA	CGGATCTCCA	2711
AGAAGATGGA	AACAATGGGC	ATCTACTTCG	CGACACCGGA	ATGGGTGGCT	CTCAACGGGC	2771
ACCGAGGCCC	AAGCCCCGGC	CAACTCAAGT	ACTGGCAAAA	CACAAGAGAA	ATACCAGAGC	2831
CCAATGAGGA	CTACCCAGAC	TATGTGCACG	CGGAGAAGAG	CCGGTTGGCG	TCAGAAGAAC	2891
AGATCCTACG	GGCAGCCACG	TCGATCTACG	GGGCTCCAGG	ACAGGCTGAA	CCACCCCAGG	2951
CCTTCATAGA	CGAGGTCGCC	AGGGTCTATG	AAATCAACCA	TGGGCGTGGT	CCAAACCAGG	3011

53

AGCAGATGAA	GGACCTGCTC	CTGACTGCGA	TGGAGATGAA	GCATCGCAAT	CCCAGGCGGG	3071
CTCCACCAAA	GCCAAAGCCA	AAACCCAATG	CTCCATCACA	GAGACCCCCT	GGACGGCTGG	3131
GCCGCTGGAT	CAGGACGGTC	TCCGACGAGG	ACTTGGAGTG	AGGCTCCTGG	GAGTCTCCCG	3191
ACACTACCCG	CGCAGGTGTG	GACACCAATT	CGGCCTTCTA	CCATCCCAAA	TTGGATCCGT	3251
TCGCGGGTCC	CCT					3264

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 145 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Asp 1 5 10 15

Gly Ser His Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala 20 25 30

Asn Asp Arg Thr Gly Val His Ser Gly Arg His Pro Gly Glu Ala His

Thr Gln Val Arg Asn Leu Asp Leu Gln Leu Asp Cys Arg Gly Tyr Arg
50 55 60

Val Arg Thr Asn Cys Leu Phe Pro Trp Ile Pro Trp Phe Ser Cys Arg
65 70 75 80

Cys Ser Leu His Thr Ala Glu Gln Trp Glu Leu Pro Ile Arg Pro Asp 85 90 95

Ala Pro Asp Ser Ala Glu Pro Ala Cys Gln Leu Gln Leu Gln Ala 100 105 110

Ser Glu Gln Glu Ser Asn Arg Thr Val Lys His Thr Pro Trp Trp Arg 115 120 125

Leu Cys Thr Lys Arg Asn His Lys Arg Ser Asp Leu Pro Arg Lys Pro 130 135 140

Glu

54

(2)	INFORMATION	FOR	SEQ	ID	NO:33:
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(i)	SEQUENCE	CHARACTER	ISTICS:
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- (A) LENGTH: 3264 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: circular
- (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 131..3169

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGA1	racg <i>i</i>	ATC	GGTC	TGAC	cc c	GGGG	GAGT	C AC	CCGG	GGAC	AGG	CCAT	CAC '	rgcc'	TTGTT	C 60
CTG	TTGO	AA	CTCC	TCTT	тс т	GCTG'	ract <i>i</i>	A TC	GTTG	ATGG	TGA	GTAG	AGA '	rcag	ACAAA	2 120
GATO	CGCAC					CTG :					Gln (169
						ATG Met 165										217
						CAC His										265
					Авр	ACA Thr										313
				Ser		GTA Val			His							361
			Gln			CAG Gln										409
						AGG Arg 245						Leu				457

														ATA Ile		505
														AGC Ser 285		553
														GGG Gly		601
GTT Val	CTA Leu	GTT Val 305	GGA Gly	GAA Glu	GGG Gly	GTG Val	ACT Thr 310	GTT Val	CTC Leu	AGT Ser	CTA Leu	CCG Pro 315	ACT Thr	TCA Ser	TAT Tyr	649
														GGA Gly		697
														AGA Arg		745
TAC Tyr	ACC Thr	ATA Ile	ACA Thr	GCT Ala 355	GCA Ala	GAT Asp	GAA Glu	TAC Tyr	CAA Gln 360	TTC Phe	TCG Ser	TCA Ser	CAA Gln	CTC Leu 365	ATC Ile	793
CCG Pro	AGT Ser	GGC Gly	GTG Val 370	AAG Lys	ACC Thr	ACA Thr	CTG Leu	TTC Phe 375	TCC Ser	GCC Ala	AAC Asn	ATC Ile	GAT Asp 380	GCT Ala	CTC Leu	841
ACC Thr	AGC Ser	TTC Phe 385	AGC Ser	GTT Val	GGT Gly	GGT Gly	GAG Glu 390	CTT Leu	GTC Val	TTC Phe	AGC Ser	CAA Gln 395	GTA Val	ACG Thr	ATC Ile	889
CAA Gln	AGC Ser 400	ATT Ile	GAA Glu	GTG Val	GAC Asp	GTC Val 405	ACC Thr	ATT Ile	CAC His	TTC Phe	ATT Ile 410	GGG Gly	TTT Phe	GAC Asp	GGG Gly	937
ACA Thr 415	GAC Asp	GTA Val	GCA Ala	GTC Val	AAG Lys 420	GCA Alá	GTT Val	GCA Ala	ACA Thr	GAC Asp 425	TTT Phe	GGG Gly	CTG Leu	ACA Thr	ACT Thr 430	985
GGG Gly	ACA Thr	AAC Asn	AAC Asn	CTT Leu 435	GTG Val	CCA Pro	TTC Phe	AAC Asn	CTG Leu 440	GTG Val	GTC Val	CCA Pro	ACA Thr	AAT Asn 445	GAG Glu	1033
ATC Ile	ACC Thr	CAG Gln	CCC Pro 450	ATC Ile	ACT Thr	TCC Ser	ATG Met	AAA Lys 455	CTA Leu	GAG Glu	GTT Val	GTG Val	ACC Thr 460	TAC Tyr	AAG Lys	1081

		GGT Gly						1129
		CAC His						1177
		TAT Tyr 500						1225
		AAC Asn						1273
		GAG Glu						1321
		CTG Leu						1369
		GAG Glu						1417
		TCA Ser 580						1465
		GCC Ala						1513
		GCT Ala						1561
		GGC Gly						1609
		AAA Lys						1657
		GAC Asp 660						1705

CAG Gln	GTG Val	CCC Pro	CAG Gln	AAT Asn 675	CCC Pro	ATT Ile	GTT Val	GAT Asp	GGC Gly 680	ATT Ile	CTG Leu	GCA Ala	TCC Ser	CCA Pro 685	GGA Gly	1753
ATC Ile	CTG Leu	CGT Arg	GGC Gly 690	GCA Ala	CAC His	AAC Asn	CTC Leu	GAC Asp 695	TGC Cys	GTG Val	CTA Leu	TGG Trp	GAG Glu 700	GGA Gly	GCC Ala	1801
ACT Thr	CTT Leu	TTC Phe 705	CCT Pro	GTT Val	GTC Val	ATT Ile	ACG Thr 710	ACA Thr	CTC Leu	GAG Glu	GAT Asp	GAG Glu 715	CTG Leu	ACC Thr	CCC Pro	1849
AAG Lys	GCA Ala 720	CTG Leu	AAC Asn	AGC Ser	AAA Lys	ATG Met 725	TTT Phe	GCT Ala	GTC Val	ATT Ile	GAA Glu 730	GGT Gly	GTG Val	CGA Arg	GAG Glu	1897
GAC Asp 735	CTC Leu	CAG Gln	CCT Pro	CCA Pro	TCC Ser 740	CAA Gln	CGG Arg	GGA Gly	TCC Ser	TTC Phe 745	ATT Ile	CGA Arg	ACT Thr	CTC Leu	TCT Ser 750	1945
GGC Gly	CAT His	AGA Arg	GTC Val	TAT Tyr 755	GGC Gly	TAT Tyr	GCC Ala	CCA Pro	GAC Asp 760	GGA Gly	GTA Val	CTG Leu	CCT Pro	CTG Leu 765	GAG Glu	1993
ACC Thr	GGG Gly	AGA Arg	GAC Asp 770	TAC Tyr	ACC Thr	GTT Val	GTC Val	CCA Pro 775	ATT Ile	GAT Asp	GAT Asp	GTG Val	TGG Trp 780	GAC Asp	GAT Asp	2041
AGC Ser	ATA Ile	ATG Met 785	CTG Leu	TCG Ser	CAG Gln	GAC Asp	CCC Pro 790	ATA Ile	CCT Pro	CCA Pro	ATC Ile	ATA Ile 795	GGG Gly	AAC Asn	AGC Ser	2089
GGC Gly	AAC Asn 800	CTA Leu	GCC Ala	ATA Ile	GCA Ala	TAC Tyr 805	ATG Met	GAT Asp	GTC Val	TTC Phe	AGG Arg 810	CCC Pro	AAG Lys	GTC Val	CCC Pro	2137
ATC Ile 815	CAC His	GTG Val	GCT Ala	ATG Met	ACA Thr 820	GGG Gly	GCC Ala	CTC Leu	AAT Asn	GCC Ala 825	CGC Arg	GGT Gly	GAG Glu	ATC Ile	GAG Glu 830	2185
AGT Ser	GTT Val	ACG Thr	TTC Phe	CGC Arg 835	AGC Ser	ACC Thr	AAA Lys	CTC Leu	GCC Ala 840	ACA Thr	GCC Ala	CAC His	CGA Arg	CTT Leu 845	GGC Gly	2233
ATG Met	AAG Lys	TTA Leu	GCT Ala 850	Gly	CCT Pro	GGA Gly	GCC Ala	TAT Tyr 855	Asp	ATT	AAT Asn	ACA Thr	GGA Gly 860	Pro	AAC Asn	2281
TGG Trp	GCA Ala	ACG Thr 865	Phe	GTC Val	AAA Lys	CGT Arg	TTC Phe 870	Pro	CAC His	AAT Asn	CCC Pro	CGA Arg 875	Asp	TGG	GAC Asp	2329

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AGG Arg	TTG Leu 880	CCC Pro	TAC Tyr	CTC Leu	AAC Asn	CTT Leu 885	CCT Pro	TAT Tyr	CTC Leu	CCA Pro	CCA Pro 890	ACA Thr	GCA Ala	GGA Gly	CGT Arg	2377
CAG Gln 895	TTC Phe	CAT His	CTA Leu	GCC Ala	CTG Leu 900	GCT Ala	GCC Ala	TCC Ser	GAG Glu	TTC Phe 905	AAA Lys	GAG Glu	ACC Thr	CCA Pro	GAA Glu 910	2425
														GAC Asp 925		2473
Leu	Phe	Arg	Ser 930	Ala	Leu	Gln	Val	Phe 935	Met	Trp	Leu	Glu	Glu 940	AAC Asn	Gly	2521
Ile	Val	Thr 945	Asp	Met	Ala	Asn	Phe 950	Ala	Leu	Ser	Asp	Pro 955	Asn	GCG Ala	His	2569
Arg	Met 960	Lys	Asn	Phe	Leu	Ala 965	Asn	Ala	Pro	Gln	Ala 970	Gly	Ser	AAG Lys	Ser	2617
														CGA Arg		2665
										qaA				TCC Ser 1005	Lys	2713
				Met					Ala					GTG Val		2761
			His					Pro					Tyr	TGG Trp		2809
		Arg					Pro					Pro		TAT Tyr		2857
	Ala					Leu					Gln			CGG Arg		2905
GCC Ala					Gly					Ala					Ala	2953

								•	_							
TTC 1 Phe :	ATA Ile	GAC Asp	GAG Glu 1090	Val	GCC Ala	AGG Arg	GTC Val	TAT Tyr 1095	Glu	ATC Ile	AAC Asn	His	GGG Gly 1100	Arg	GGT Gly	3001
CCA I	AAC Asn	CAG Gln 1105	Glu	CAG Gln	ATG Met	AAG Lys	GAC Asp 1110	Leu	CTC Leu	CTG Leu	ACT Thr	GCG Ala 1115	Met	GAG Glu	ATG Met	3049
Lys 1	CAT His 1120	Arg	AAT Asn	CCC Pro	AGG Arg	CGG Arg 1125	Ala	CCA Pro	CCA Pro	AAG Lys	CCA Pro 1130	Lys	CCA Pro	AAA Lys	CCC Pro	3097
AAT (Asn)	Ala	CCA Pro	TCA Ser	CA G Gln	AGA Arg 1140	Pro	CCT Pro	GGA Gly	CGG Arg	CTG Leu 1145	Gly	CGC Arg	TGG Trp	ATC Ile	AGG Arg 1150	3145
			GAC Asp		Asp			TGAG	GCT(CT C	egga(TCT(CC C	GACA(CTACC	3199
CGCGCAGGTG TGGACACCAA TTCGGCCTTC TACCATCCCA AATTGGATCC GTTCGCGGGT 3												3259				
cccc	T															3264
(2) INFORMATION FOR SEQ ID NO:34: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1013 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear																
			MOLE													
	•							: SE								
Met 1	Thr	Asn	Leu	Met 5	Asp	His	Thr	Gln	Gln 10	Ile	Val	Pro	Phe	Ile 15	Arg	
Ser	Leu	Leu	Met 20	Pro	Thr	Thr	Gly	Pro 25	Ala	Ser	Ile	Pro	Asp 30	Asp	Thr	
Leu	Glu	Lys 35		Thr	Leu	Arg	Ser 40		Thr	Ser	Thr	Tyr 45	Asn	Leu	Thr	
Val	Gly 50		Thr	Gly	Ser	Gly 55		lle	Val	Phe	Phe 60	Pro	Gly	Phe	e Pro	
Gly	Ser	Val	Val	Gly	Ala		Tyr	Thr	Leu	Gln 75	Ser	Ser	: Gly	/ Asr	Tyr 80	

Gln Phe Asp Gln Met Leu Leu Thr Ala Gln Asn Leu Pro Ala Ser Tyr

				85					90					95	
Asn	Tyr	Сув	Arg 100	Leu	Val	Ser	Arg	Ser 105	Leu	Thr	Val	Arg	Ser 110	Ser	Thr
Leu	Pro	Gly 115	Gly	Val	Tyr	Ala	Leu 120	Asn	Gly	Thr	Ile	Asn 125	Ala	Val	Thr
Phe	His 130	Gly	Ser	Leu	Ser	Glu 135	Leu	Thr	Asp	Tyr	Ser 140	Tyr	Asn	Gly	Leu
Met 145	Ser	Ala	Thr	Ala	Asn 150	Ile	Asn	Азр	Lys	Ile 155	Gly	Asn	Val	Leu	Val 160
Gly	Glu	Gly	Val	Thr 165	Val	Leu	Ser	Leu	Pro 170	Thr	Ser	Tyr	Asp	Leu 175	Ser
Tyr	Val	Arg	Leu 180	Gly	Asp	Pro	Ile	Pro 185	Ala	Ala	Gly	Leu	Asp 190	Pro	Lys
Leu	Met	Ala 195	Thr	Сув	Asp	Ser	Ser 200	Asp	Arg	Pro	Arg	Val 205	Tyr	Thr	Ile
Thr	Ala 210	Ala	Asp	Glu	Tyr	Gln 215	Phe	Ser	Ser	Gln	Leu 220	Ile	Pro	Ser	Gly
Val 225	Lys	Thr	Thr	Leu	Phe 230	Ser	Ala	Asn	Ile	Asp 235	Ala	Leu	Thr	Ser	Phe 240
Ser	Val	Gly	Gly	Glu 245	Leu	Val	Phe	Ser	Gln 250	Val	Thr	Ile	Gln	Ser 255	Ile
Glu	Val	Asp	Val 260	Thr	Ile	His	Phe	Ile 265	Gly	Phe	Asp	Gly	Thr 270	Asp	Val
Ala	Val	Lys 275	Ala	Val	Ala	Thr	Asp 280	Phe	Gly	Leu	Thr	Thr 285	Gly	Thr	Asn
Asn	Leu 290	Val	Pro	Phe	Asn	Leu 295	Val	Val	Pro	Thr	Asn 300	Glu	Ile	Thr	Gln
Pro 305	Ile	Thr	Ser	Met	Lys 310	Leu	Glu	Val	Val	Thr 315	Tyr	Lys	Ile	Gly	Gly 320
Thr	Ala	Gly	Asp	Pro 325	Ile	Ser	Trp	Thr	Val 330	Ser	Gly	Thr	Leu	Ala 335	Val
Thr	Val	His	Gly 340	Gly	Asn	Tyr	Pro	Gly 345	Ala	Leu	Arg	Pro	Val 350	Thr	Leu
Val	Ala	Tyr 355	Glu	Arg	Val	Ala	Ala 360	Gly	Ser	Val	Val	Thr 365	Val	Ala	Gly

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Val	Ser 370	Asn	Phe	Glu	Leu	Ile 375	Pro	Asn	Pro	Glu	Leu 380	Ala	Lys	Asn	Leu
Val 385	Thr	Glu	Tyr	Gly	Arg 390	Phe	Asp	Pro	Gly	Ala 395	Met	Asn	Tyr	Thr	Lys 400
Leu	Ile	Leu	Ser	Glu 405	Arg	Asp	Arg	Leu	Gly 410	Ile	Lys	Thr	Val	Trp 415	Pro
Thr	Arg	Glu	Tyr 420	Thr	Asp	Phe	Arg	Glu 425	Tyr	Phe	Met	Glu	Val 430	Ala	Asp
		435					440					445	Lys		
	450					455					460		Thr		
Pro 465	Pro	Ala	Ala	Pro	Leu 470	Ala	His	Ala	Ile	Gly 475	Glu	Gly	Val	Asp	Tyr 480
Leu	Leu	Gly	Asp	Glu 485	Ala	Gln	Ala	Ala	Ser 490	Gly	Thr	Ala	Arg	Ala 495	Ala
Ser	Gly	Lys	Ala 500	Arg	Ala	Ala	Ser	Gly 505	Arg	Ile	Arg	Gln	Leu 510	Thr	Leu
Ala	Ala	Asp 515	Lys	Gly	Cys	Glu	Val 520	Val	Ala	Asn	Met	Phe 525	Gln	Val	Pro
Gln	Asn 530	Pro	Ile	Val	Asp	Gly 535	Ile	Leu	Ala	Ser	Pro 540	Gly	Ile	Leu	Arg
545					550					555			Thr		560
				565					570				Lys	575	
		_	580					585					Asp 590		
		595		-			600					605	Gly		
	610		-			615					620		Thr		
Asp 625	Tyr	Thr	Val	Val	Pro 630	Ile	Asp	Asp	Val	Trp 635	Asp	Asp	Ser	Ile	Met 640

- Leu Ser Gln Asp Pro Ile Pro Pro Ile Ile Gly Asn Ser Gly Asn Leu 645 650 655
- Ala Ile Ala Tyr Met Asp Val Phe Arg Pro Lys Val Pro Ile His Val 660 665 670
- Ala Met Thr Gly Ala Leu Asn Ala Arg Gly Glu Ile Glu Ser Val Thr 675 680 685
- Phe Arg Ser Thr Lys Leu Ala Thr Ala His Arg Leu Gly Met Lys Leu 690 695 700
- Ala Gly Pro Gly Ala Tyr Asp Ile Asn Thr Gly Pro Asn Trp Ala Thr 705 710 715 720
- Phe Val Lys Arg Phe Pro His Asn Pro Arg Asp Trp Asp Arg Leu Pro 725 730 735
- Tyr Leu Asn Leu Pro Tyr Leu Pro Pro Thr Ala Gly Arg Gln Phe His
 740 745 750
- Leu Ala Leu Ala Ala Ser Glu Phe Lys Glu Thr Pro Glu Leu Glu Asp
 755 760 765
- Ala Val Arg Ala Met Asp Ala Ala Ala Asn Ala Asp Pro Leu Phe Arg 770 775 780
- Ser Ala Leu Gln Val Phe Met Trp Leu Glu Glu Asn Gly Ile Val Thr 785 790 795 800
- Asp Met Ala Asn Phe Ala Leu Ser Asp Pro Asn Ala His Arg Met Lys 805 810 815
- Asn Phe Leu Ala Asn Ala Pro Gln Ala Gly Ser Lys Ser Gln Arg Ala 820 825 830
- Lys Tyr Gly Thr Ala Gly Tyr Gly Val Glu Ala Arg Gly Pro Thr Pro 835 840 845
- Glu Glu Ala Gln Arg Glu Lys Asp Thr Arg Ile Ser Lys Lys Met Glu 850 855 860
- Thr Met Gly Ile Tyr Phe Ala Thr Pro Glu Trp Val Ala Leu Asn Gly 865 870 875 880
- His Arg Gly Pro Ser Pro Gly Gln Leu Lys Tyr Trp Gln Asn Thr Arg 885 890 895
- Glu Ile Pro Glu Pro Asn Glu Asp Tyr Pro Asp Tyr Val His Ala Glu 900 905 910

- Lys Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser 915 920 925
- Ile Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp 930 935 940
- Glu Val Ala Arg Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln 945 950 955 960
- Glu Gln Met Lys Asp Leu Leu Leu Thr Ala Met Glu Met Lys His Arg 965 970 975
- Asn Pro Arg Arg Ala Pro Pro Lys Pro Lys Pro Lys Pro Asn Ala Pro 980 985 990
- Ser Gln Arg Pro Pro Gly Arg Leu Gly Arg Trp Ile Arg Thr Val Ser 995 1000 1005
- Asp Glu Asp Leu Glu 1010

Claims

1. A method for preparing live Birnavirus, comprising the following steps:

preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts, transfecting host cells with said synthetic RNA transcripts, incubating said host cells in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 2. The method according to claim 1, wherein said Birnavirus is infectious bursal disease virus.
- 3. The method according to claim 1, wherein said host cells are African green monkey Vero cells.
- 4. The method according to claim 1, wherein said segments A and B of said cDNA are independently prepared.
- 5. The method according to claim 4, wherein said segment A is present in plasmid pUC19FLAD78 or pUC18FLA23.
- 6. The method according to claim 4, wherein said segment B is present in plasmid pUC18FLBP2.
- 7. A live infectious bursal disease virus, wherein said virus is made by a process comprising the steps of preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce a synthetic RNA transcript, transfecting a host cell with said synthetic RNA transcript, incubating said host cell in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 8. A synthetic RNA encoding proteins VP1, VP2, VP3, VP4, and VP5 of infectious bursal disease virus.
 - 9. A host cell transfected with the synthetic RNA according to claim 8.
- 10. A cDNA containing at least a portion of the infectious bursal disease virus genome selected from the group consisting of segment A,

segment B and segments A and B of infectious bursal disease virus, wherein said cDNA includes the 5' and 3' terminii of said segments.

- 11. A recombinant vector comprising the cDNA according to claim 10.
- 12. The vector according to claim 11, wherein said vector is a plasmid.
- 13. The vector according to claim 12, wherein said plasmid is selected from the group consisting of pUC19FLAD78, pUC18FLA23 and pUC19FLBP2.
 - 14. A host cell transformed with the vector according to claim 11.
- 15. A vaccine comprising an infectious bursal disease virus according to claim 7, wherein said infectious bursal disease virus is inactivated or attenuated prior to administration.
- 16. A method for producing a live infectious bursal disease virus vaccine, comprising the steps of

preparing a full-length cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts, purifying said synthetic RNA transcripts,

transfecting host cells with said purified RNA transcripts,

incubating said host cells in a culture medium,

isolating live infectious bursal disease virus from said culture medium, attenuating said live infectious bursal disease virus to produce a virus with reduced virulence, and

combining said live infectious bursal disease virus with a pharmaceutically acceptable carrier to produce a live infectious bursal disease virus vaccine.

- 17. The method according to claim 16, wherein said live infectious bursal disease virus is attenuated by serial passage or site directed mutagenesis.
- 18. The method according to claim 1, wherein said host cells are poultry cells.
- 19. The method according to claim 18, wherein said poultry cells are chicken, turkey, or quail cells.

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20. The method according to claim 19, wherein said poultry cells are chicken embryo fibroblast cells or chicken embryo kidney cells.

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Fig. 1

Fig. 4

Fig. IA

Fig. IB

Fig. IC

Fig. 4A

Fig.4B

Fig. 5A

Fig. 5B

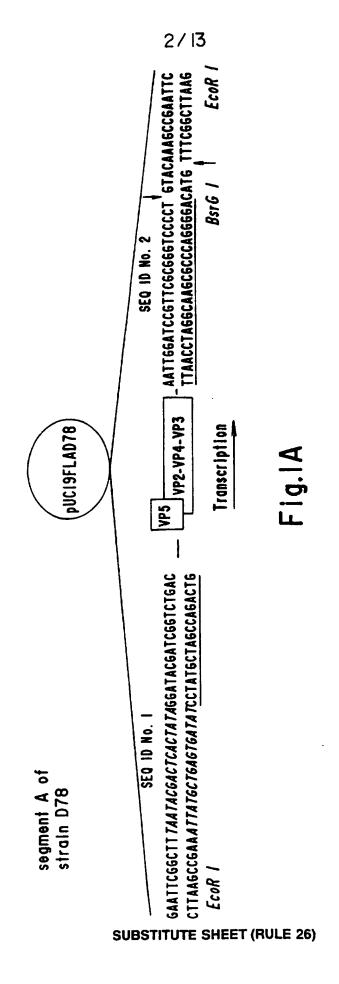
Fig. 6

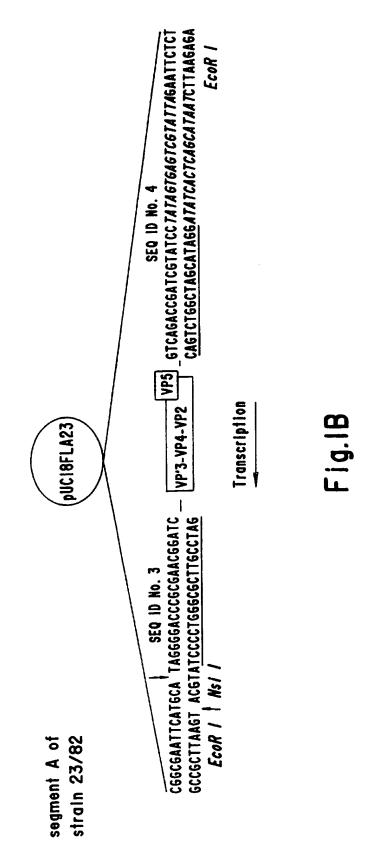
Fig. 5

Fig. 6A

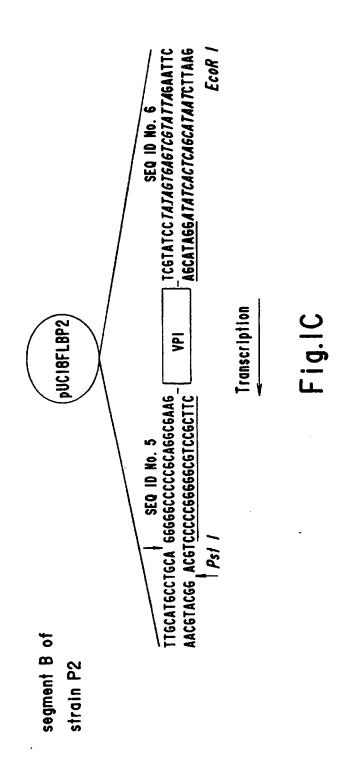
Fig. 6B

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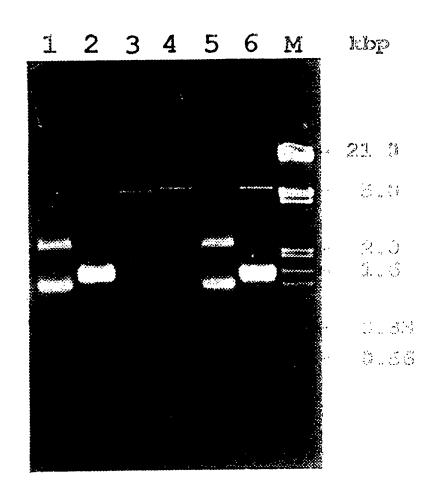


Fig. 2

Fig.3

	530	540	550	260	570	280
23-82A	GEAAGCCTGAGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTGCGAAC	6AGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTG	TACAGCTACA	ACGGCTGAT	FTCAGCCACT	GCGAAC
23A/P2B	66AAGCCTGAGTGAGTTGACTGACTACAGCTACAGGGCTGATGTCAGCCACTGCGAAC	AGTTGACTGAC	TACAGCTACA	ACGGGCTGAT	GTCAGCCACT	GCGAAC
SEQ 10 No. 8 P2A SEQ 10 No. 9	66AAGCCTGAGTGACTGATGTTAGCTACATGGTTGATGTCTGCAACAGCCAAC 530 540 550 560 570 580	AACTGACAGAT 540	GTTAGCTACA 550	ATGGGTTGA1 560	retctecaca 570	GCCAAC 580
23-82A SFO 10 NO. 7	590 600 610 620 630 640 ATCAACGACAAGGGAAGGGGTGACTGTTCAGTCTACCG	600 TCGGGAACGTT	610 CTAGTTGGA	620 SAAGGGTGA(630 CTGTTCTCAGI	640 CTACCG
23A/P2B SE0 ID No. 8	ATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAAGGGGTGACTGTTCTCAGTCTACCG	TCGGGAACGT	CTAGTT66A	SAGGGGTGA	CTETTCTCAGI	CTACCE
P2A SEQ ID No. 9	ATCAACGACAAATTGGGAACGTCCTAGTAGGGGAAGGGGTCACCGTCCTCAGCTTACCC 590 630 640	TTGGGAACGTC 600	CTAGTAGGG	5AA6666TCA(620	CCTCCTCAGG 630	TTACCC 640

Segment A

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		130	140	150	091	170	081
23-82B SFO ID No 10	9	TTTCAATAGTCCACAGGCGCGAACGAAGATCTCAGCAGCGTTCGGCATAAAGCCTACTG	ACAGGGGGAA	CGAAGATCT	CAGCAGCETT	CECATAAAG	CTACTE
23A/P2B	: :	TITICAACAGICCACAGGCGCGAAGCACGATCTCAGCAGCGTTCGGCATAAAGCCTACTG	ACAGGGGGAA	GCACGATCT	TCTCAGCAGCGTTCGGC	CECATAAEC	CTACTE
SEQ 10 NO. 11 P28 SEQ 10 No. 12	= 2	TITICAACAGTCCACAGGGGGGGATCTCAGCAGGGTTCGGCATAAAGCCTACTG	CCACAGGCGCGAAGCACCATCTCAGCAGGGTTCGGCAT	GCACGATCT 150	CAGCAGCETT	SECATAAGC 170	CTACT6 180
23-828	9	190 200 210 220 230 240 CTGGACAAGACTCTTGATCCCCAAAGTCTGGGGTGCCACCTGAGGATCCGC	200 Ggagaa ctct	210 TGATCCCCA	220 AAGTCT666T	230 SCCACCTGAGG	240 ATCC6C
23A/P2B	≥ =	CTGGACAAGACGTGGAAGACTCTTGATCCCTAAAGTTTGGGTGCCACCTGAGGATCCGC	GEAGAACTCT	TGATCCCTA	AAGTTTGGGT	SCCACCTGAGE	ATCCGC
P28 SEQ ID No. 12	- 2	CTGGACAAGACGTGGAAGTCTTGATCCCTAAAGTTTGGGTGCCACCTGAGGATCCGC	GGAAGACTCT 200	TGATCCCTA 210	AAGTTTGGGT 220	SCCACCTGAGG 230	ATCCGC 240

Fig.3E

GGGAGTACTTCATGGAGGTTGCAGATCTCAACTCACCCCTAAAGATTGCAGGAGCATTTGGCTTTAAGGA CTAATACTGAGTGAGAGAGATCGTCTAGGCATCAAGACAGTCTGGCCCACCAGGGAGTACACCGATTTGA CATAATCCGAGCCATTCGGAAGATTGCGGTGCCAGTGGTATCCACACTCTTCCCTCCAGCTGCACCCCTA GAGCCGCGTCAGGAAAGCTAGAGCTGCCTCAGGACGAATAAGGCAGCTAACTCTCGCAGCTGACAAGGG GTGCGAGGTAGTCGCCAACATGTTCCAGGTGCCCCAGAATCCCATTGTTGATGGCATTCTGGCATCCCCA CCGAAGTTGATGGCCACGTGCGACAGTAGTGACAGACCCAGAGTCTACACCATAACAGCTGCAGATGAAT CACCAGCTTCAGCGTTGGTGGTGAGCTTGTCTTCAGCCAAGTAACGATCCAAAGCATTGAAGTGGACGTC **ACCATTCACTTCATTGGGTTTGACGGGACAGACGTAGCAGTCAAGGCAGTTGCAACAGACTTTGGGCTGA** CAACTGGGACAACAACCTTGTGCCATTCAACCTGGTGGTCCCAACAAATGAGATCACCCAGCCCATCAC TCCATGAAACTAGAGGTTGTGACCTACAAGATTGGCGGCACCGCTGGTGACCCAATATCATGGACAGTG **AGTGGTACACTAGCTGTGACGGTGCACGGAGGCAACTACCCTGGGGCTCTCCGTCCTGTCACCCTGGTGG** CCTATGAACGAGTGGCTGCAGGATCTGTTGTCACAGTTGCAGGGGTGAGCAACTTCGAGCTAATCCCCAA CCCTGAGCTTGCAAAGAACCTAGTTACAGAGTATGGCCGCTTTGACCCCGGAGCAATGAACTACACCAAA **GCACATGCAATCGGAGAGGTGTAGACTACCTCCTGGGCGACGAGGCCCAAGCAGCCTCAGGGACAGCTC** CATTCCGGACGACACCCTGGAGAAGCACACACTCAGGTCCGAAACCTCGACTTACAACTTGACTGTAGGG GATACAGGGTCAGGACTAATTGTCTTTTTCCCTGGATTCCCTGGTTCAGTTGTAGGTGCTCACTACACAC CTACT6CA66CTAGT6A6CA66A6TCTAACCGTAC6GTCAA6CACACTCCCT6GT6GCGTTTAT6CACTA **GGATACGATCGGTCTGACCCCGGGGAGTCACCCGGGGACAGGCCATCACTGCCTTGTTCCTGGTTGGAA** IGATGTCAGCCACTGCGAACATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAAGGGGTGACTGTTCT CTCCTCTTTCTGCTGTACTATCGTTGATGGTGGTAGAGATCAGACAAACGATCGCAGCGATGACAAACC | GATGGATCACACCCAACAGATTGTTCCGTTCATACGGAGCCTTCTGATGCCAACGACCGGACCGGCGTC **ACCAATTCTCGTCACAACTCATCCCGAGTGGCGTGAAGACCACACTGTTCTCCGCCAACATCGATGCTCT** 491 631 771 841 981 331 471 051 261 211 281 281 351 421 <u>6</u>

 $\overline{2}$

TCCATCTAGCCCTGGCTGCCTCC GAGTTCAAAGAGACCCCAGAACTCGAAGACGCTGTGCGCG CAATGG AT6CC6CT6CAAAT6CC6ACCCATTGTTCC6CTCA6CTCTCCA6GTCTTCATGTGGTTGGAAGAAACG6 **GATTGTGACCGACATGGCTAACTTCGCCCTCAGCGACCCAAACGCGCATAGGATGAAAAACTTCCTAGCA** 5AGGCCCCACACCAGAAGAGGCACAGAGGGAAAAAGACACACGGATCTCCAAGAAGATGGAAACAATGGG CATCTACTTC6C6ACACCG6AATG6GTG6CTCTCAACGG6CACCGAGGCCCAAGCCCCGGCCAA CTCAAG actegcaaaacacaagagaataccagagcccaatgagactacccagactatgtegacgcgggagga SCCGGTTGGCGTCAGAAGAACAGATCCTACGGGCAGCCACGTCGATCTACGGGGCTCCAGGACAGGCTGA **ACCACCCC AGGCCTTCATAGACGAGGTCGCCAGGGTCTATGAAATCAACCATGGGCGTGGTCCAAACCAG** SAGCAGATGAAGGACCTGCTCCTGACTGCATGGAGATGAAGCATCGCAATCCCAGGCGGGGTCCACCAA 16CCAAAGCCAAAACCCAATGCTCCATCACAGAGCCCCCTGGACGGCTGGGCCGCTGGATCAG GACGGT TCCGACG AGGACTTGGAGTGAGGCTCCTGGGAGTCTCCCGACACTACCCGCGCGGGTGT GGACACCAAT GGCTATGCCCCAGACGGAGTACTGCCTCTGGAGACCGGGAGAGACTACACCGTTGTCCCAATTGATGATG CATAGCATACATGGATGTCTTCAGGCCCAAGGTCCCCATCCACGTGGCTATGACAGGGGCCCTCAATGCC AGCTGGTCCTGGAGCCTATGACATTAATACAGGACCTAACTGGGCAACGTTCGTCAAACGTTTCCCTCA **GTGGGACGATAGCATAATGCTGTCGCAGGACCCCATACCTCCAATCATAGGGAACAGCGGCAACCTAGC** CGCGGTGAGATCGAGAGTGTTACGTTCCGVAGCACCAAACTCGCCACAGCCCACGGACTTGGCATGAAGT ITACGACACTCGAGGATGAGCTGACCCCCAAGGCACTGAACAGCAAAATGTTTGCTGTCATTGAAGGTGT SCGAGAGGACCTCCAGCCTCCATCCCAACGGGGATCCTTCATTCGAACTCTCTGGGCCATAGAGTCTA1 rcgg ccttctaccatcccaaattggatccgttcgcgggtccct 2451 2521 2591 2661 2801 2871 2941 3081 2311 2381 2731 301 2241 2031 2101 2171

Total number of bases is: 3264. DNA sequence composition: 834 A; 942 C; 853 G; 635

Sequence name: 23-82A (SEQ ID NOS: 31 and 33)

Fig.4B

TGCAGGGCAATGGGAACTACAAGTTCGATCAGATGCTCCTGACTGCCCAGAACCTACCGGCCAGTTACAA CTACTGCAGGCTAGTGAGTCGGAGTCTCACAGTGAGGTCAAGCACATTCCTGGTGGCGTTTATGCACTA CCGCCACCGACAACCTTATGCCATTCAATCTTGTGATTCCAACAAACGAGATAACCCAGCCAATCACATC **ACGAAAGAGTGGCAACAGGATCCGTCGTTACGGTCGCTGGGGTGACCAACTTCGAGCTGATCCCAAATCC** GAACTAGCAAAGAACCTGGTTACAGAATACGGCCGATTTGACCCCAGGAGCCATGAACTACACAAAATTG AATCCGGGCCATAAGGAGGATAGCTGTGCCGGTGGTCTCCACATTGTTCCCACCTGCCGCTCCCCTAGCC CATGCAATTGGGGAAGGTGTAGACTACCTGCTGGGCGATGAGGCACAGGCTGCTTCAGGAACTGCTCGAG CCCCTCAGGAAAAGCAAGAGCTGCCTCAGGCCGCATAAGGCAGCTGACTCTCGCCGCCGACAAGGGGTA CGAGGTAGTCGCGAATCTATTCCAGGTGCCCCAGAATCCCGTAGTCGACGGGATTCTTGCTTCACCTGGG STACTCCGCGGTGCACACACCTCGACTGCGTGTTAAGAGAGGGTGCCACGCTATTCCCTGTGGTTATTA SATTCCGGACGACACCCTGGAGAAGCACACTCTCAGGTCAGAGACCTCGACCTACAATTTGACTGTGGG CAGCTTACCCACATCATATGATCTTGGGTATGTGAGGCTTGGTGACCCCATTCCCGCAATAGGGCTTGAC CCAAAAATGGTAGCCACATGTGACAGCAGTGACAGGCCCAGAGTCTACACCCATAACTGCAGCCGATGATT **AATACTTCATGGAGGTGGCCGACCTCAACTCTCCCCTGAAGATTGCAGGAGCATTCGGCTTCAAAGACAT** CTCCTCCTTCTACAACGCTATCATTGATGGTTAGTAGAGATCAGACAAACGATCGCAGCGATGACAAACC SACACAGGGTCAGGGCTAATTGTCTTTTTCCCTGGATTCCCTGGCTCAATTGTGGGTGCTCACTACACAC **ATCTACCTCATAGGCTTTGATGGGACAACGGTAATCACCAGGGCTGTGGCCGCAAACAATGGGCTGACGA 366A6CCTAGCAGTGACGATCCATGGTGGCAACTATCCAGGGGCCCTCCGTCCCGTCACGCTAGTGGCCT AACGGCACCATAAACGCCGTGACCTTCCAAGGAAGCCTGAGTGAACTGACAGATGTTAGCTACAATGGGT** IGATGTCTGCAACAGCCAACATCAACGACAAAATTGGGAACGTCCTAGTAGGGGAAGGGGTCACCGTCCT ACCAATTCTCATCACAGTACCAACCAGGTGGGGTAACAATCACACTGTTCTCAGCCAACATTGATGCCA CACAAGCCTCAGCGTTGGGGGAGAGCTCGTGTTTCAAACAAGCGTCCACGGCCTTGTACTGGGCGCCACC ATACTGAGTGAGGGGGCCGTCTTGGCATCAAGACCGTCTGGCCAACAAGGGGAGTACACTGACTTTCGT GCAAGATCAAACCCAACAGATTGTTCCGTTCATACGGAGCCTTCTGATGCCAACAACGGACCGGCGT 20 561 631 701 771 841 981 981 <u>6</u> 261 331 04 211 281 351 421 491 47 54

Fig.5A

CAGATGAAAGATCTGCTCTTGACTGCGATGGAGATGAAGCATCGCAATCCCAGGCGGGCTCTACCAAAGC CACCTTGCCATGGCTGCATCAGAGTTCAAAGAGACCCCCGAACTCGAGAGTGCCGTCAGAGCAATGGAAG **CAGCAGCCAACGTGGACCCACTATTCCAATCTGCACTCAGTGTTCATGTGGCTGGAAGAGAATGGGAT ACCCCAAGCTTTCATAGACGAAGTTGCCAAAGTCTATGAAATCAACCATGGACGTGGCCCAAACCAAGAA** CCAAGCCAAAACCCAATGCTCCAACACAGAGACCCCTGGTCGGCTGGGCCGCTGGATCAGGACCGTCTC **GGCGAGATTGAGAAAGTAAGCTTTAGAAGCACCAAGCTCGCCACTGCACACCGACTTGGCCTTAGGTTGG CTGGTCCCGGAGCATTCGATGTAAACACCGGGCCCAACTGGGCAACGTTCATCAAACGTTTCCCTCACAA TCCACGCCACTGGGACAGGCTCCC CTACCTCAACCTACCATACCTTCCACCCAATGCAGGACGCCAGTAC** GTGACTGACATGGCCAACTTCGCACTCAGCGACCCGAACGCCCATCGGATGCGAAATTTTCTTGCAAAC **GCACCACAAGCAGCAGCAAGTCGCAAAGGGCCAAGTACGGGACAGCAGGCTACGGAGTGGAGGCTCGGG GCCCCACACCAGAGGAAGCACAGAGGGAAAAAGACACACGGATCTCAAAGAAGATGGAGACCATGGGCAT** FGATGAGGACCTTGAGTGAGGCTCCTGGGAGTCTCCCGACACCACCCCGCGCGGGTGTGGACACCAATTCG **CGACAGTGGAAGACGCCATGACACCCAAAGCATTGAACAGCAAAATGTTTGCTGTCATTGAAGGCGTGCG** TATGCTCCAGATGGGGTACTTCCACTGGAGACTGGGAGAGACTACACCGTTGTCCCAATAGATGATGTCT CTACTTTGCAACACCCAGAATGGGTAGCACTCAATGGGCACCGAGGGCCAAGCCCCGGCCAGCTAAAGTAC **GGTTGGCATCAGAAGAACAAATCCTAAGGGCAGCTACGTCGATCTACGGGGCTCCAGGACAGGCAGAGGC AGCTTACATGGATGTGTTTCGACCCAAAGTCCCATTCCATGTGGCTATGACGGGAGCCCTCAATGCTTGT GGGACGACACCATTATGCTGTCCAAAGATCCCATACCTCCTATTGTGGGAAACAGTGGAAATCTAGCCA** SCCTTACAACATCCCAAATTGGATCCGTTCGCGGGTCCCCT 2801 252| 259| 266| 2731 2871 294 30 3081 2241 2311 2381 2451 2031 2101 2171

Total number of bases is: 3261.

DNA sequence composition: 873 A; 909 C; 847 G; 632 T; 0 0THER;

Sequence name: D78F (SEQ ID NOS: 27 and 29)

Fig.5B

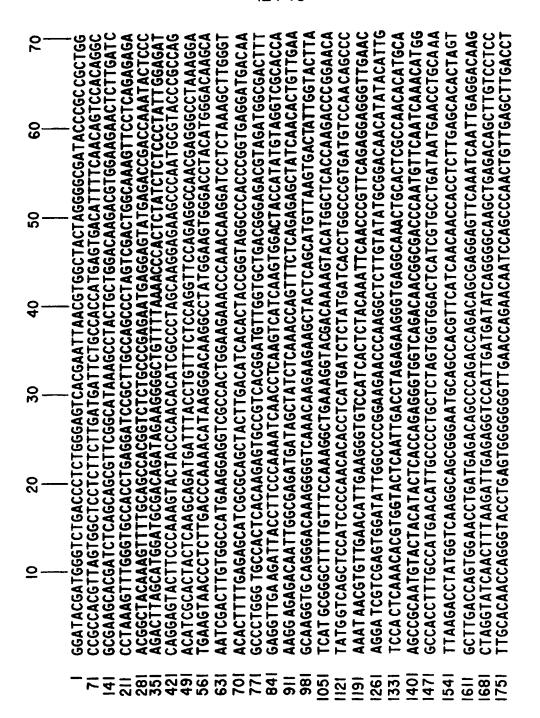


Fig.6A

TGTTGGGCTCCACCTGCCCGCCAAGAGAGCCACCGGTGTCCAGGCCGCTCTTCTCGGAGCAGGAACGAG CAGACCAATGGGGATGGAGGCCCCAACACGGTCCAAGAACGCCGTGAAAATGGCCAAACGGGGGGAACGC CAAAAGGAGGCCGCTAACAGCCATGATGGGAACCACTCAAGAAGAGGACACTAATCCCAGACCCGGTAT CTCGTCCTTCTAGCCACAGCAAGAAGCCGTCTGCAAGATGCAGTTAAGGCCAAGGCAGAAGCCGAGAAAC **GGAGAAAGCCGACATCGCCAGCAAGGTCGCCCACTCAGCACTCGTGGAAACAAGCGACGCCCTTGAAGCA** STTCAGTCGACTTCCGTGTACACCCCCAAGTACCCAGAAGTCAAGAACCCACAGACCGCCTCCAACCCCG CTGAGAGCCTAGCCGAACTGAACAAGCCAGTACCCCCCAAGCCCCCAAATGTCAACAGACCAGTCAACAC TTTGTTCTGCTGCGTATCCCAAGGGAGTAGAGAACAAGAGTCTCAAGTCCAAAGTCGGGATCGAGCAGG **CTGCAAGAATAACGCAGGCGCCGCTCGGCGCATCTGGAGGCCCAAGGGGTTCCCACTCGACGAGTTCCTA GCCGAGTGGTCTGAGCTGTCAGAGTTCGGTGAGGCCTTCGAAGGCTTCAATATCAAGCTGACCGTAACAT** TCCACAAGTCCAAGCCAGACGACCCCGATGCAGACTGGTTCGAAAGATCAGAACTCTGTCAGACCTTCT CATACAAGGTAGTCAGGTATGAGGCGTTGAGGTTGGTAGGTGGTTGGAACTACCCACTCCTGAACAAAGC <u> actagggtggtcagctacatacagcaaagatctcgggatctatgtgccggtgcttgacaaggaacgcct</u> SCCCGGCCTTCGCCTGCGGGGGCCCCC 2591 2381 2451 2521 2661 2241 2031 2101 2171 2311

Total number of bases is: 2827.

DNA sequence composition: 796 A; 770 C; 724 G; 537 T; 0 OTHER;

Sequence name: P2B (SEQ ID No: 25)

Fig.6B

INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

	SSIFICATION OF SUBJECT MATTER :Please See Extra Sheet.			
US CL	:Please See Extra Sheet. to International Patent Classification (IPC) or to both	national classification and IPC		
	LDS SEARCHED			
	documentation searched (classification system followed	d by classification symbols)		
	424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236,			
Documenta	tion searched other than minimum documentation to the	extent that such documents are included	in the fields searched	
	data base consulted during the international search (na N-MEDLINE, BIOSIS, CAPLUS, CABA	me of data base and, where practicable	e, search terms used)	
C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.	
х	MUNDT et al. Complete Nucleotide Noncoding Regions of Both Genome So of Infectious Bursal Disease Virus. Viro 10-18, see entire document.	egments of Different Strains	1-2, 4-20	
x	US 4,530,831 A (LUTTICKEN ET AL see entire document.	.) 23 JULY 1985 (07/23/85),	7, 15-20	
X	US 5,192,539 A (VAN DER MAREL (09/03/93), see entire document.	. ET AL) 09 MARCH 1993	1-3, 7, 15-20	
x	MUNDT et al. Identification of a nove bursal disease virus-infected cells. Jon 1995, Vol. 76, pages 437-443, see enti	urnal of General Virology.	8	
X Further documents are listed in the continuation of Box C. See patent family annex.				
Special categories of cited documents:				
"A" do	cument defining the general state of the art which is not considered be of particular relevance	data and not in conflict with the appl the principle or theory underlying the		
		"X" document of particular relevance; the considered novel or cannot be consider		
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the publication date of another citation or other cited to establish the				
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	cument published prior to the international filing date but leter than a priority date claimed	"&" document member of the same patent	(family	
Date of the	actual completion of the international search	Date of mailing of the international ser	arch report	
22 SEPTI	EMBER 1997	1 0 NOV 1997		
Commission Box PCT	Washington, D.C. 20231			

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12955

C (Continu	tion). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the releva	ant passages	Relevant to claim No
x	BAYLISS et al. A comparison of the sequences of segm four infectious bursal disease virus strain and identificat variable region in VP2. Journal of General Virology. 19 71, pages 1303-1312, see entire document.	ion of a	1-2, 5-8, 10-13
Y	MORGAN et al. Sequence of the Small Double-Strander Genomic Segment of Infectious Bursal Disease Virus and Deduced 90kDa Product. Virology. 1988, Vol. 163, page see entire document.	d Its	1-20
Y	SPIES et al. Nucleotide sequence of infectious bursal disgenome segment A delineates two major open reading fi Nucleic Acids Research. 1989, Vol. 17, No. 19, page 79 entire document.	rames.	1-20
Y	WO 91/16925 A1 (UNIVERSITY OF MARYLAND at COLLEGE PARK) 14 NOVEMBER 1991 (14/11/91), se document.		1-20
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

	A. CLASSIFICATION OF SUBJECT MATTER: IPC (6):
	A61K 39/00, 39/38, 39/12; C12P 21/04; C12N 7/00, 7/01, 7/02, 7/04, 7/06, 7/08, 15/00, 15/09, 15/63, 15/70, 15/74
	A. CLASSIFICATION OF SUBJECT MATTER: US CL :
	424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236, 237, 238, 239, 320.1; 536/23.72
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